Chapter 8 Climate-Smart Agriculture Practices for Mitigating Greenhouse Gas Emissions



M. Zaman, K. Kleineidam, L. Bakken, J. Berendt, C. Bracken,
K. Butterbach-Bahl, Z. Cai, S. X. Chang, T. Clough, K. Dawar, W. X. Ding,
P. Dörsch, M. dos Reis Martins, C. Eckhardt, S. Fiedler, T. Frosch, J. Goopy,
C.-M. Görres, A. Gupta, S. Henjes, M. E. G. Hofmann, M. A. Horn,
M. R. Jahangir, A. Jansen-Willems, K. Lenhart, L. Heng,
D. Lewicka-Szczebak, G. Lucic, L. Merbold, J. Mohn, L. Molstad, G. Moser,
P. Murphy, A. Sanz-Cobena, M. Šimek, S. Urquiaga, R. Well,
N. Wrage-Mönnig, S. Zaman, J. Zhang, and C. Müller

Abstract Agricultural lands make up approximately 37% of the global land surface, and agriculture is a significant source of greenhouse gas (GHG) emissions, including carbon dioxide (CO₂), methane (CH₄) and nitrous oxide (N₂O). Those GHGs are

M. Zaman $(\boxtimes) \cdot L$. Heng

e-mail: m.zaman@iaea.org; zamanm_99@yahoo.com

K. Kleineidam · C. Eckhardt · A. Jansen-Willems · G. Moser · C. Müller Institute of Plant Ecology, Justus Liebig University Giessen, Giessen, Germany

L. Bakken Norwegian University of Life Sciences (NMBU), Aas, Norway

J. Berendt · S. Fiedler · N. Wrage-Mönnig University of Rostock, Rostock, Germany

C. Bracken

School of Agriculture and Food Science and Earth Institute, University College Dublin, Dublin, Ireland

K. Butterbach-Bahl Institute of Meteorology and Climate Research, Atmospheric Environmental Research (IMK-IFU), Karlsruhe Institute of Technology, Karlsruhe, Germany

Z. Cai

School of Geography Sciences, Nanjing Normal University, Jiangsu, China

S. X. Chang

Department of Renewable Resources, University of Alberta, Edmonton, AB T6G 2E3, Canada

T. Clough Department of Soil & Physical Sciences, Faculty of Agriculture & Life Sciences, Lincoln University, Lincoln, New Zealand

© The Author(s) 2021 M. Zaman et al. (eds.), *Measuring Emission of Agricultural Greenhouse Gases and Developing Mitigation Options using Nuclear and Related Techniques*, https://doi.org/10.1007/978-3-030-55396-8_8

Soil and Water Management & Crop Nutrition (SWMCN) Section, Joint FAO/IAEA Division of Nuclear Techniques in Food and Agriculture, International Atomic Energy Agency (IAEA), Vienna, Austria

responsible for the majority of the anthropogenic global warming effect. Agricultural GHG emissions are associated with agricultural soil management (e.g. tillage), use of both synthetic and organic fertilisers, livestock management, burning of fossil fuel for agricultural operations, and burning of agricultural residues and land use change. When natural ecosystems such as grasslands are converted to agricultural production, 20-40% of the soil organic carbon (SOC) is lost over time, following cultivation. We thus need to develop management practices that can maintain or even increase SOC storage in and reduce GHG emissions from agricultural ecosystems. We need to design systematic approaches and agricultural strategies that can ensure sustainable food production under predicted climate change scenarios, approaches that are being called climate-smart agriculture (CSA). Climate-smart agricultural management practices, including conservation tillage, use of cover crops and biochar application to agricultural fields, and strategic application of synthetic and organic fertilisers have been considered a way to reduce GHG emission from agriculture. Agricultural management practices can be improved to decreasing disturbance to the soil by decreasing the frequency and extent of cultivation as a way to minimise soil C loss and/or to increase soil C storage. Fertiliser nitrogen (N) use efficiency can be improved to reduce fertilizer N application and N loss. Management measures can also be taken to minimise agricultural biomass burning. This chapter reviews the current literature on CSA practices that are available to reduce GHG emissions and

K. Dawar

Department of Soil and environmental Sciences, University of Agriculture, Peshawar, Pakistan

W. X. Ding Institute of Soil Science, Chinese Academy of Sciences, Nanjing, China

P. Dörsch · L. Molstad Faculty of Environmental Sciences and Natural Resource Management, Norwegian University of Life Sciences (NMBU), Aas, Norway

T. Frosch

Leibniz Institute of Photonic Technology, Technical University Darmstadt, Darmstadt, Germany

J. Goopy International Livestock Research Institute (ILRI), Nairobi, Kenya

C.-M. Görres

Department of Soil Science and Plant Nutrition/Department of Applied Ecology, Hochschule Geisenheim University, Geisenheim, Germany

A. Gupta Independent Consultant India, Mumbai, India

S. Henjes · M. A. Horn Institute of Microbiology, Leibniz University Hannover, Hannover, Germany

M. E. G. Hofmann Picarro B.V., 's-Hertogenbosch, The Netherlands

M. M. R. Jahangir Department of Soil Science, Bangladesh Agricultural University, Mymensingh, Bangladesh increase soil C sequestration and develops a guideline on best management practices to reduce GHG emissions, increase C sequestration, and enhance crop productivity in agricultural production systems.

Keywords Agriculture \cdot Carbon dioxide \cdot Climate-smart agriculture \cdot C sequestration \cdot GHG \cdot Methane \cdot Mitigation \cdot Nitrous oxide

8.1 Introduction on Climate-Smart Agriculture Practices and Greenhouse Gas Emissions

Agriculture is a major source of greenhouse gases (GHGs) that affect climate change and is also itself a victim of climate change. Agricultural lands make up 37.6% of the global land surface, and agriculture is a significant source of GHG emissions (IPCC 2014; Smith et al. 2008), where CO₂, CH₄ and N₂O are the major forms of trace gases that are responsible for the majority of the global warming effect. Agricultural GHG emissions are associated with agricultural soil management (e.g. tillage), use of both synthetic and organic fertilisers, livestock management, burning of fossil fuel for agricultural operations and burning of agricultural residues. In particular, agriculture can be the source for 52% and 84%

D. Lewicka-Szczebak

G. Lucic Picarro Inc., Santa Clara, CA, USA

L. Merbold Mazingira Centre, International Livestock Research Institute (ILRI), Nairobi, Kenya

J. Mohn

Laboratory for Air Pollution & Environmental Technology, Empa Dübendorf, Dübendorf, Switzerland

P. Murphy

Environment & Sustainable Resource Management Section, School of Agriculture & Food Science, and UCD Earth Institute, University College, Dublin, Ireland

A. Sanz-Cobena

Research Center for the Management of Environmental and Agricultural Risks (CEIGRAM), ETSIAAB, Universidad Politécnica de Madrid, Madrid, Spain

M. Šimek

Institute of Soil Biology, Biology Centre of the Czech Academy of Sciences, and Faculty of Science, University of South Bohemia, České Budějovice, Czech Republic

M. dos Reis Martins · S. Urquiaga

EMBRAPA Agrobiologia Seropédica, Brazilian Agricultural Research Corporation, Seropedica, RJ, Brazil

R. Well

Thünen Institute of Climate-Smart Agriculture, Braunschweig, Germany

Laboratory of Isotope Geology and Geoecology, Institute of Geological Sciences, University of Wrocław, Wrocław, Poland

of global anthropogenic emissions of CH₄ and N₂O, respectively (Smith et al. 2008). Since the global warming potentials of CH₄ and N₂O are much higher than that of CO₂ based on per unit mass and a 100-year time frame (IPCC 2014), advanced concepts are required to reduce agricultural emissions of CH₄ and N₂O.

In addition to causing increased levels of GHG emission, human settlement in previously unpopulated areas means that natural ecosystems are converted to agricultural production, with 20-40% of the SOC lost following cultivation and with most of that loss occurring in the first a few years (Davidson and Ackerman 1993). A recent estimate indicates that 133 billion tonnes of SOC, which is about 8% of the total global SOC stock, had been lost from the top 2 metres of soil on a global scale since agriculture started about 12,000 years ago, with the rate of loss dramatically increased since the industrial revolution (Sanderman et al. 2017). The Sanderman et al. (2017) study also indicated that the percentage of SOC loss was greater on cropland but the total amount of SOC loss was slightly higher on grazing land as more than twice as much land is grazed. This indicates that there is a greater potential to improve the % SOC gain in cropland but there is a greater potential to increase total SOC storage in grazing land. One of the key aspects of SOC is that the soil and vegetation stores about three times the organic C of the atmosphere (Plate 8.1; FAO 2004), and thus small changes in the organic C stock in the soil and vegetation can cause a large effect on atmospheric CO₂ concentration; therefore, great efforts must be made to increase SOC storage in and to reduce GHG emissions from terrestrial ecosystems. In managed systems, SOC storage can be increased by management practices such as avoiding the burning of crop residues after harvest, and the application of composts and biochar and animal manure to increase organic C input to the soil.

There is significant potential for the agriculture sector to contribute to the reduction of anthropogenic sources of GHG emissions since agriculture is a large source of GHG and to increase soil C storage since much soil C has been lost through cultivation (Plate 8.2; top panel). For example, agricultural management practices can be improved to reduce disturbance to the soil by decreasing the frequency and extent of cultivation as a way to minimise soil C loss and/or to increase soil C storage; if permanent vegetation can be maintained, soil C storage can increase, benefiting from the C cycle becoming more closed in the system and the soil being able to trap more C (Plate 8.2; bottom panel). Fertiliser N use efficiency (NUE) can be improved through strategic application of fertiliser so as to reduce N loss, whether it is through

S. Zaman

J. Zhang

K. Lenhart

Bingen University of Applied Sciences, Berlinstr. 109, Bingen 55411, Germany

University of Canterbury, Christchurch, New Zealand

School of Geography, Nanjing Normal University, Nanjing, China



Plate 8.1 An illustration of the distribution of carbon in terrestrial ecosystems, including the atmosphere. (*Source* Schwartz 2014)

leaching or gaseous form of N loss. Management measures can also be taken to minimise agricultural biomass burning. Climate-smart agriculture (CSA) management practices, including strategic use of synthetic and organic fertilisers and water, conservation tillage, use of cover crops, and the use of amendments such as biochar, nitrification inhibitors and lime to agricultural fields, have been considered a way to reduce GHG emissions from agriculture (Bai et al. 2019; Lipper et al. 2014; Zaman et al. 2008a and 2009; Zaman and Blennerhassett 2010). The FAO defines CSA as a systematic approach for developing agricultural strategies that can ensure sustainable food security under predicted climate change scenarios (FAO 2013). Based on this definition, a range of agricultural practices can be developed to help improve food security and environmental quality simultaneously in the context of global change.



Plate 8.2 An illustration of a disturbance to the soil causes an increased release of CO_2 and other greenhouse gases (and leading to the soil being leaky, open treasure chest) and in soils with undisturbed vegetation will cause the carbon cycle to be more closed (*Source* FAO 2013)

Since the soil can act as a sink or source for CO_2 and affect climate change, if we can enhance the C sink strength and remove more CO_2 from the atmosphere by adopting CSA, then we can be in a win–win situation by not only combating the negative effects of climate change but also improving soil quality and health, including the retention of nutrients and water, and increased agricultural productivity.

8.2 Climate-Smart Agricultural Technology to Reduce GHG Emissions

Climate-smart agriculture emphasises on improving risk management, enhancing information flows and promoting local institutions to increase the adaptive capacity of communities to climate change (Campbell et al. 2014), as such CSA plays a pivotal role in maintainable development. Climate-smart agricultural practices such as the use of cover crops, amendments and tillage management play key roles in reducing agricultural GHG emissions. Take N₂O, a potent GHG, as an example, the emission of N₂O is affected by many factors, such as the use of N process inhibitors (urease and nitrification inhibitors), the strategic application of synthetic fertilisers (the right type, the right amount that is based on crop requirement and soil tests, at the right plant growth stage, and the right method (e.g. even spread)), avoiding the application of N fertilisers to very wet or very dry soils, maintaining soil pH above

6 by adding lime to lower the $N_2O:N_2$ ratio, co-application of animal manure and chemical fertilisers to provide a more balanced nutrient supply, optimising animal stocking rate to avoid over-grazing, keeping animals off the pasture especially in the wet season to minimise N input and avoid soil compaction, and minimising the excessive use of farm machinery. Many of those fall under CSA practices. Below, key CSA practices and their effect on GHG emissions from agriculture are discussed in detail.

8.2.1 Nitrogen Process Inhibitors and Greenhouse Gas Emissions

Multiple microbial soil processes contribute to GHG emissions. One of the major processes contributing to GHG emissions is the mineralization of organic matter by microbial organisms, a process also called soil heterotrophic respiration, where organic C is converted to CO_2 and released to the atmosphere. Nitrification and denitrification are key processes contributing to N₂O emissions from the soil, where ammonia-oxidation and successive nitrifier denitrification or denitrification, among other processes, can be important pathways for N₂O emissions (Fig. 7.8) (Guo et al. 2018). In the ammonia oxidation process, N₂O can be produced by the chemical decomposition of hydroxylamine (NH₂OH). One potential way to mitigate N₂O emissions is to use nitrification and urease inhibitors to slow down the rate of nitrification and reduce the availability of the substrate (NH₄⁺) for nitrification.

8.2.1.1 Synthetic Nitrification Inhibitors

Nitrification is the process where ammonium or ammonia is converted to nitrate, via nitrite as an intermediate N species. Since the formation of nitrite is usually the rate-limiting step in the nitrification process, nitrite usually does not accumulate in the soil, unless the soil has a high pH where nitrite oxidisers are inhibited (Rodgers 1986). During the nitrification process, N₂O can be formed and emitted from the soil. The final product of nitrification, i.e nitrate, is subject to leaching losses, and when anaerobic conditions develop, nitrate is denitrified thus leading to the formation of N₂O. Both nitrate leaching and denitrification can both conserve N in the soil and reduce N₂O emissions (Abalos et al. 2012). In this respect, the application of nitrification inhibitors can effectively reduce nitrification rates and the buildup of nitrate which can further be denitrified. Nitrification inhibitors are a group of chemical compounds that slow down the conversion rate of ammonium or ammonia to nitrate by inhibiting the ammonia monooxygenase activity, through disrupting the activity of *Nitrosomonas* and *Nitrobacter* bacteria (Abalos et al. 2014). Nitrification

inhibitors are applied with ammonium-, ammonia- or urea-based fertilisers as the application of such fertilisers substantially increases the rate of nitrification. The need for nitrification inhibitors in non-fertilised soils is rare as nitrification rates in such soils are low. The ammonium or ammonia can come from urine, manures, composts or crop residues as they decompose, or fertilisers such as ammonium sulphate or urea (Rodgers 1986). Under suitable conditions (e.g. warm soil temperature and a moisture content near field capacity, when there is still ample O_2 available), nitrification occurs within days or weeks of the application of ammonium-based fertilisers and thus if nitrification can be effectively reduced within that timeframe, N loss from the system can be substantially reduced (Sanz-Cobena et al. 2017).

There are at least eight compounds commercially available as nitrification inhibitors but the most commonly used ones are 2-chloro-6-(trichloromethyl)pyridine (nitrapyrin), dicyandiamide (DCD), 3,4-dimethylpyrazole phosphate (DMPP, e.g. ENTEC[®]) (IPNI undated) and pronitradine (CenturoTM). Nitrapyrin (commercial product: N-ServeTM and InstinctTM) can be applied to the soil in different ways: if anhydrous ammonia is the fertiliser being applied, the nitrification inhibitor can be injected with the fertiliser, if solid N fertilisers are being applied, the inhibitor can be coated onto the fertiliser, and if manure is used as an organic fertiliser, the inhibitor can be mixed with the manure before manure application. Nitrapyrin is usually effective for less than 30 days after being applied to the soil and is volatile, therefore, the best way to apply this inhibitor is to incorporate it into the soil; DMPP can be effective for reducing nitrification rates for 25–70 days and is usually preblended with fertilisers; DCD can last 25-55 days and can be coated on solid fertilisers (Sanz-Cobena et al. 2012), or surface applied to soils that have been applied with manure or on grazing land to reduce nitrification from urine patches (IPNI undated). While DMPP is somewhat immobile, DCD can be relatively easily leached from the soil; those behaviours of the nitrification inhibitors should be considered when they are applied in the field.

DCD and DMPP have been found to be equally effective in changing soil inorganic N content, leaching of dissolved inorganic N (DIN) and emissions of N_2O in a recent meta-analysis of field trials reported in the literature (Yang et al. 2016). Their costbenefit analysis (CBA) showed that the economic benefit was about seven times greater with DCD than with DMPP when applied with ammonium-based fertilisers to reduce nitrification (Yang et al. 2016). Those two nitrification inhibitors are among the most commonly used. DCD is cheaper and less volatile, but the application rate of DMPP is typically one-tenth that of DCD, and DMPP has a lower eco-toxicological effect for plants as is summarised in Yang et al. (2016). In an Australian study, DMPP application (as ENTEC®) decreased N_2O emissions by 15% in a subtropical pasture in Queensland (Lam et al. 2018).

The effectiveness of the nitrification inhibitors can be affected by soil properties such as soil water content (Barrena et al. 2017), and soil organic matter and clay content (Zhu et al. 2019). Zhu et al. (2019) reported that the efficiency of DMPP in reducing nitrification and N_2O emissions was lower in soils with high organic matter

and clay contents, likely due to the high rates of adsorption of DMPP by soil organic matter and clay. The effectiveness of DCD in reducing N₂O emissions from urine patches in New Zealand is highly season-specific, with reductions of 52, 39 and 16% in autumn, spring and summer, respectively, but DCD application increased NH₃ emissions by 56, 9 and 17% in the respective seasons (Zaman et al. 2009). Management practices can also affect the efficiency of nitrification inhibitors. For example, biochar application to the soil has been shown to decrease the efficiency of DMPP both at 40 and 80% of water-filled pore space (WFPS) in a laboratory incubation study (Fuertes-Mendizábal et al. 2019). The use of nitrification inhibitors increases crop yield and NUE, but the effectiveness was greatest when they are used in coarse-textured soils, irrigated systems and/or crops receiving high rates of N fertiliser input (Abalos et al. 2014).

8.2.1.2 Synthetic Urease Inhibitors

Urease inhibitors retard the activity of urease, which exists in the soil and plant residues. Urease is involved in the conversion of urea to ammonium in a process called hydrolysis. Unfortunately, the urea hydrolysis process increases the pH of the soil and causes a large proportion of the formed ammonium to be volatilized as ammonia. Urease inhibitors would slow down the rate of hydrolysis or the rate of release of ammonium, reduce the loss of N as ammonia through volatilization, and increase the NUE of urea fertilisers applied to the soil. One of the main reasons for the improved NUE is for more urea to be washed into the soil over time (when the rate of hydrolysis is suppressed), as urea is highly soluble in water. The better contact with the soil increases the chances of the released ammonium to be adsorbed by the cation exchange sites.

The N-(n-butyl) thiophosphoric triamide (NBPT) and N-(n-propyl) thiophosphoric triamide (NPPT) are two chemicals that have been shown to be effective in inhibiting urease activities (Rodriguez et al. 2019; Lam et al. 2018; Zaman et al. 2009; Sanz-Cobena et al. 2008). Products containing those urease inhibitors include AgrotainTM (that contains NBPT) and LimusTM (that contains both NBPT and NPPT). The application of NBPT prior to urine deposition was more effective in reducing ammonia volatilization loss (17.5–27.6% reduction) as compared with applying the NBPT after the urine deposition (0.6–2.9% reduction) in pastureland in New Zealand (Rodriguez et al. 2019). The effectiveness of NBPT is highly season-specific for reducing NH₃ volatilization loss from urine patches in New Zealand, with reductions of 29, 93 and 31% in autumn, spring and summer, respectively (Zaman et al. 2009). An Australian study reached similar conclusions that urea applied with NBPT (as Green UreaNV®) was effective in decreasing NH₃ volatilization (by 44%) in a subtropical pasture in Queensland (Lam et al. 2018). The effectiveness of NBPT was greater in alkaline soils (pH \geq 8) (Abalos et al. 2014). It has also been observed, under laboratory conditions, that application of urease inhibitors in soils where nitrification is the main pathway in the production of N_2O (i.e. WFPS < 50%), could be an effective way to mitigate these N losses (Sanz-Cobena et al. 2014).

The combined application of urease inhibitor + the nitrification inhibitor, DCD, (e.g. Agrotain Plus) to inhibit both the hydrolysis of urea and the nitrification processes to minimise the N loss showed best results in maximising NUE. When urea and ammonium nitrate (UAN) was applied at 150 mg N kg⁻¹ in a sandy loam soil in the United States with Agrotain Plus, N₂O emissions were reduced by 78% compared to the control (Cai et al. 2018). The use of double inhibitors containing both NBPT and DCD (3:7) in a New Zealand study reduced NH₃ volatilization by 14, 78 and 9% in autumn, spring and summer, respectively, and N₂O emissions by 37, 67 and 28%, respectively, from urine patches (Zaman et al. 2009). However, another study showed that adding DCD with NBPT did not further reduce NH₃ volatilization loss, but in fact enhanced the volatilization loss by maintaining a higher soil NH₄⁺ concentration and pH for a longer period of time after urea application, indicating that DCD co-applied with NBPT and urea could offset the effect of NBPT in reducing volatilization losses (Soares et al. 2012).

8.2.1.3 Biological Nitrification Inhibitors

Some plant species can release secondary metabolites through root exudation and/or from leaf litter. Such metabolites have the ability to suppress microbial nitrification (Souri and Neumann 2018). In some earlier studies, detectable biological nitrification inhibition (BNI) was found in root exudates of sorghum (Sorghum bicolor (L.)), pearl millet (Pennisetum glaucum (L.) R. Br.) and groundnut (Arachis hypogaea (L.)) among tested cereal and legume crops, while *Brachiaria humidicola* (Rendle) Schweick and B. decumbens Stapf had the highest BNI capacity among pasture grass species tested (Subbarao et al. 2007). In addition, when BNI compounds from root exudates were applied to the soil, their inhibitory effects on NO_3^- formation lasted for more than 50 days (Subbarao et al. 2007). Linoleic acid, α -linolenic acid, and methyl linoleate, a fatty acid methyl ester of linoleic acid, are some of the example compounds that are effective in biologically inhibiting nitrification (Subbarao et al. 2008). Subbarao et al. (2007) suggested that some level of BNI is likely a widespread phenomenon in tropical pasture grass species and those properties could be used to suppress nitrification in natural or managed systems. The production in and release from plants of BNIs are triggered by the presence of NH_4^+ in the rhizosphere of plants, which means that BNIs are released where the majority of the nitrifier populations reside (Subbarao et al. 2013a). The pH in the rhizosphere will affect the release of BNIs from roots; for example, sorghum plants release BNIs from their roots in the presence of NH_4^+ when the rhizosphere pH is between 5.0 and 6.0 (Subbarao et al. 2013b), indicating the usefulness of BNI to reduce nitrification in alkaline soils will be non-existent or very low. More research is needed to understand and take advantage of BNI in agricultural production systems to reduce N loss and improve NUE.

8.2.2 Soil Amendments and Greenhouse Gas Emissions

8.2.2.1 Mulch

Addition of mulch to the soil will change the availability of carbon (C) and other nutrients to microbial populations and will thus affect soil GHG emissions. Existence of a litter layer (mulch) can induce microbial N immobilisation in the litter layer, and result in reduced available N and reduced plant growth rate; however, the litter layer may benefit plant growth by conserving soil moisture (Matsushima and Chang 2006) but may reduce soil temperature and N mineralization rates (Matsushima and Chang 2007). Addition of mulch can immobilise mineral N in the soil and reduce the availability of NH₄⁺ for nitrification and NO₃⁻ for denitrification, and thus reduce N₂O emissions as compared to no mulch addition (Wu et al. 2013). Using wood bark mulch reduced the nitrate concentration in the soil and cut soil N₂O emissions by up to 28% in a grape (Vitus vinifera L. cv. Merlot) yard on a sandy loam soil in British Columbia, Canada, when measured over a two-year period (Fentabil et al. 2016). However, overloading of straw to the soil surface can delay seed germination and result in the need for additional fertiliser supply to compensate for the N that may be immobilised in the critical period of the early growing season (Procházková et al. 2003). As far as CO_2 emissions are concerned, mulching will usually result in increased CO_2 emissions due to the addition of labile C in the mulch, with the rate of CO_2 emissions increasing with the increased rate of mulch addition (Wu et al. 2013). Major anthropogenic sources of methane emissions include fossil fuel production, landfills and livestock farming, but some agricultural soils can be an anthropogenic source of methane emissions as well (IPCC 2007). In rice paddy systems, straw application has been shown to increase CH₄ emissions (Bossio et al. 1999; Ma et al. 2008); however, straw addition significantly reduced CH₄ emissions under an aerobic condition in a laboratory incubation experiment, indicating that under upland conditions, straw application increased the soil's ability to take up CH₄ (Tate et al. 2007).

8.2.2.2 Biochar

Biochar has been widely studied for its effects on GHG emissions. Biochar addition to the soil can change a range of soil properties, including the cycling of C and N. Biochar application has been widely reported to reduce N₂O emissions (Wu et al. 2013; Cayuela et al. 2013; Chang et al. 2016; Hüppi et al. 2016). The application of biochar to the soil has been shown to reduce denitrification and decrease N₂O emissions by 10–90% when tested on 14 different agricultural soils, where a consistent reduction of the N₂O/(N₂ + N₂O) ratio was observed, indicating that biochar reduces N₂O emissions by facilitating the last step of the denitrification process and producing more N₂ rather than N₂O (Cayuela et al. 2013). However, in some soils,

biochar application can stimulate nitrification and increase N_2O emissions; therefore, the effect of biochar application on N_2O emissions is related to the dominant N_2O formation pathway that operates in a soil (Sánchez-García et al. 2014).

Biochar application has been reported to reduce CH_4 emissions from paddy soils, one of the largest anthropogenic sources of CH_4 emissions on a global scale (Feng et al. 2012). The reduction in paddy CH_4 emissions by biochar application was not a result of the inhibition of methanogenic archaea, but resulted from increased methanotrophic proteobacterial abundances and decreased ratios of methanogenic to methanotrophic abundances (Feng et al. 2012).

In agricultural production systems, a large quantity of crop residues are produced and the return of crop residues in the raw form vs after the crop residue is converted to biochar can have dramatic effects on the emissions of all three trace gases (Wu et al. 2013), and most research suggests that there are substantial beneficial effects to be gained in mitigating climate change by converting crop residues to biochar and applying biochar to the soil instead. The effect of biochar on GHG emissions itself is highly complex as many factors alter the function of biochar on C and N transformation processes and thus GHG emissions. When the effect of the stable C input in the form of biochar is considered, the application of biochar is considered an effective technique to mitigate climate change due to its negative emissions potential (0.7 Gt Ceq. Yr^{-1}) and its lower impact on land, water use, albedo, energy requirement and cost as compared to other negative emissions technologies such as direct air capture, increased weathering that takes up CO₂ from the air, bioenergy projects with C capture and storage, and afforestation/deforestation (Smith 2016).

8.2.2.3 Liming to Shift the Balance Between N₂O and N₂ Emissions

Soil pH is one of the key regulators of microbiological processes that affect N₂O and N₂ production and their ratio. The soil pH threshold for nitrification is 5; however, nitrification can occur even below pH 5 as some nitrifier strains are adapted to acidic conditions (Bouwman 1990). Denitrification has been reported to occur over a wide range of soil pH (5–8) (Flessa et al. 1998); however, laboratory experiments with artificially adjusted soil pH suggest that, under optimised conditions (very low pO₂, NO_3^- and with glucose amendment), denitrification can proceed even at pHs below 4 or above 10 (Šimek et al. 2002). Numerous laboratory and field studies have shown that soil pH affects N₂O and N₂ emissions and thus the ratio of these gases (e.g. Stevens and Laughlin 1998). In experiments conducted under controlled environmental conditions, raising soil pH to 7 through lime application has been found to significantly increase N₂ emissions from pasture and wetland soils treated with cow urine, urea and KNO₃ at 200 kg N ha⁻¹ (Zaman et al. 2007, 2008b). Similar trends of enhanced N₂ emissions after raising soil pH to 7 was observed in pasture soils treated with urea/urine in a field experiment (Zaman and Nguyen 2010). In another study, a site with the greatest animal impact, the ratio of N_2 to N_2O produced during denitrifying enzyme activity (DEA) measurements was fivefold higher, and the pH

was 2 units higher, than a site with the least animal impact, indicating that soil conditions were favourable for production of N_2 rather than N_2O in the area with intense excretal returns and treading (Hynšt et al. 2007).

Most researchers attribute high N_2O and low N_2 emissions in acidic conditions to the suppression of N_2O -reductase (inhibition starts at soil pH 4.5) (Daum and Schenk 1998; Flessa et al. 1998; Stevens and Laughlin 1998; Zaman et al. 2007). It is also possible that denitrifying enzymes are susceptible at low soil pH and produce N_2O from intermediate products (Nägele and Conrad 1990). However, the lower rates of N_2 emissions and higher $N_2O:N_2$ ratio at low soil pH could be due to lower amounts of soil organic C and mineral N available to the denitrifying population rather than a direct effect of low pH on denitrification enzymes (Šimek and Cooper 2002). Regardless of the biochemical mechanism for soil pH effects on N_2 emissions, raising soil pH through the application of amendments such as lime appears a viable approach to mitigate N_2O emissions (Šimek et al. 2002; Zaman and Nguyen 2010; Zaman et al. 2007, 2008b).

8.2.3 Fertiliser Type and Management and Greenhouse Gas Emissions

The use of different fertiliser types and the management of fertiliser applications can have marked effects on nitrification, denitrification and GHG emission rates (Mosier et al. 2006; Wang et al. 2018). As discussed in the earlier section, the type of fertiliser (some fertilisers are acid-forming while others raise soil pH when applied to the soil) applied can affect the total amount of GHG emitted as well as the N₂O:N ratio in the emissions. A number of agronomic practices have been widely tested to minimise N losses from agricultural production systems, for example, alteration of the rate or timing of fertiliser applications, such as autumn vs. spring, basal vs. broadcast, deep vs. surface applications, point injection placement of solutions, foliar applications of urea (Subbarao et al. 2013b), and split application. Fertilisers such as polythene-coated urea (PCU) that releases N slowly in the soil have been demonstrated to reduce nitrification (Zvomuya et al. 2003); however, the use of such fertilisers can be limited by the high cost for purchasing such fertilisers (Subbarao et al. 2013b).

8.2.4 Cropping Systems and Greenhouse Gas Emissions

8.2.4.1 Agroecosystems

The type of cropping system used has a significant effect on GHG emissions as cropping systems will differ in their fertilisation regime, crop productivity (and thus the amount of organic matter input to and retention in the soil), crop species being used (N-fixing vs. non-N-fixing crop species), water management practices, and tillage management, among others (Snyder et al. 2009). On an extreme case, GHG emission rates and timing will be dramatically different between rice cultivation in flooded fields vs. wheat production in well-drained upland sites in a rice-wheat rotation, where the rice-wheat belt makes up 24–27 million ha in South and East Asia (Wassmann et al. 2004).

Rice cultivation in paddy fields is a unique cropping system in Southeast Asia. In Vietnam, for example, rice production is the largest source of agricultural GHG, with 37.4 Tg CO₂ equivalent of total emissions, that account for 58% of agricultural GHG emissions in that country (United Nations 2013). Many factors, such as the management of fertiliser applications, animal manure and crop residue management, water regime used during rice production, and use of urease and/or nitrification inhibitors, will affect the emissions of GHGs from paddy fields. One of the biggest concerns of GHG emissions from paddy fields is the emission of CH₄ as paddy fields are mostly submerged in water during the growing season and anaerobic condition caused substantial CH₄ emission to occur. The CH₄ emissions are the balance of CH_4 production and oxidation in the soil, and are affected by factors influencing the transportation of CH₄ from the anoxic soil/free-standing water to the atmosphere (Aulakh et al. 2001). Up to 80% of the CH_4 produced in paddy soils is oxidised in the rhizosphere or microsites that are less anaerobic before it is released to the atmosphere (Sass et al. 1991; Holzapfel-Pschorn et al. 1985) and this process helps to dramatically reduce the rate of CH₄ emissions from paddy soils.

Proper water management can substantially reduce CH₄ emissions from paddy fields; midseason drainage has been reported to reduce CH₄ emissions by 44% and alternating wetting-drying cycles at ten-day intervals by 61% as compared to continuously flooded management in southeast China (Lu et al. 2000). Application of urea can increase CH₄ emissions from paddy fields as ammonium can inhibit CH₄ oxidation (Conrad and Rothfuss 1991). However, others found that application of ammonium-based fertilisers can reduce CH₄ emissions as ammonium enhances methanotrophic bacteria activities in the rhizosphere of rice plants (Bodelier et al. 2000). The effect of ammonium on CH₄ oxidation is dependent on the CH₄ concentration: inhibition at low initial CH₄ concentration (500 μ l l⁻¹) but stimulation at high initial CH₄ concentration (2000 µ11⁻¹) (Cai and Mosier 2000). However, ammonium sulphate has been found to be a promising fertiliser to use, as opposed to urea, to mitigate CH₄ emissions as sulphate enhances sulphate-reducing bacteria activities, which decrease the availability of substrates for methanogens in submerged soils (Yagi et al. 1997). Reductions in CH_4 emissions after ammonium sulphate application has been reported to range from 10 to 67% (Schütz et al. 1989; Wassmann et al. 2000). Ammonium is the preferred N form for rice, therefore, applying ammonium form of N that does not enhance CH₄ emissions would be beneficial for the environment. Application of sulphate-containing phosphorus fertilisers (Achtnich et al. (1995) and gypsum (CaSO₄) (Lindau und Bollich (1993)), a common soil amendment for sodic and/or alkaline soil reclamation, have been shown to reduce CH₄ emissions

from paddy soils. Therefore, choice of fertiliser is important in minimising CH_4 emissions from rice paddies, in this case, the use of sulphate-based N or phosphorus fertilisers would be preferred in flooded rice fields.

Applying organic fertilisers such as animal manure and returning crop residue to the soil are necessary to maintain and enhance the sustainability of rice production. However, addition of organic materials to rice fields would increase CH_4 emissions, and organic materials application to the soil can lower the soil redox potential and supply C to methanogens that are responsible for CH₄ production in paddy soils. The production of CH₄ in paddy soils is markedly influenced by the quality and quantity of organic materials added to the soil (Minasny et al. 2017). Therefore, proper manure and crop residue management strategies need to be developed to achieve environment-friendly rice cultivation. One potential alternative to the application of crop residues to paddy soil is to convert crop residues to biochar that is slow to decompose after soil application (Ippolito et al. 2012); biochar application to soil can increase soil aeration and soil C content but mitigate CH₄ emissions as compared with the conventional crop residue application (Feng et al. 2012; Karhu et al. 2011; Liu et al. 2011; Xie et al. 2013). However, other studies reported increased CH_4 emissions from rice fields after biochar application, which may be related to increased substrate supply and enhanced environment for methanogenic activity, and increased rice growth (Knoblauch et al. 2011; Lehmann et al. 2011). The effects of biochar application on CH₄ emissions is thus dependent on soil type, agricultural management practices used, and the type of biochar applied (Lehmann et al. 2011; Waters et al. 2011). Site-specific research should be conducted before any recommendation on agricultural management practices is made for farmers to adopt.

In rice paddies, the flooded condition is conducive for denitrification to occur. Even though less attention has been paid to N_2O as compared to CH_4 emissions from paddy fields, N loss in the form of NH_3 volatilisation and N_2O emissions and NO_3^- leaching affect GHG emissions as well as NUE in rice production systems. Therefore, urease inhibitors can be used to slow down the rate of urea hydrolysis when urea is used as the main N fertiliser (Rogers et al. 2015), while the use of nitrification inhibitors can reduce nitrate leaching loss (Li et al. 2008) and N_2O emissions (Majumdar et al. 2000; Kumar et al. 2000).

8.2.4.2 Organic Farming

Organic farming has long been considered a viable agricultural practice to improve soil health, reduce the resource use intensity, reduce the environmental impact of agriculture and improve food quality (Squalli and Adamkiewicz 2018). The comprehensive study of Squalli and Adamkiewicz (2018), based on longitudinal state-level data in the United States collected between 1997 and 2010, demonstrates that a 1% increase in organic farming acreage can result in a 0.049% reduction in GHG emissions; however, they showed that the net effect of organic farming on GHG emissions is dependent on the contribution of transportation (fuel burning) on methane and nitrous oxide emissions, even though their calculation indicates that the negative environmental effect of transportation output associated with organic food production is small relative to the environmental benefits of organic farming. A study in Switzerland showed a 40% reduction in N₂O emissions for organic compared to conventional systems; however, yield-scaled N₂O emissions under silage maize were not different between organic and conventional systems (Skinner et al. 2019), indicating that even if we consider the lower yield in organic systems, organic farming does not have a negative effect on GHG emissions. In contrast, Aguilera et al. (2015) and Cayuela et al. (2017) showed in two Meta-analyses that fertilisation with solid organic manures (the most used form of N fertilisers in organic agroecosystems) led to the lowest N₂O emission factor in Mediterranean cropping systems worldwide. On the other hand, we must recognise that organic farming does have a lower crop yield (c. 25% on average) which may require a larger area of land to be cultivated to produce the same amount of food compared to conventional farming (Kniss et al. 2016).

8.2.4.3 Row, Intercropping and Crop Rotation

When dealing with a cropping system that involves rice production (e.g. in Southeast Asia), it is often difficult to strike a balance in the mitigation of different GHGs. Using a rice-wheat crop rotation system as an example, the two crops markedly differ in their nature and intensity of GHG fluxes, where CH_4 emissions are a major contributor to GHG emissions from rice paddies. In rice production systems, water regimes, rice cultivars and soil properties all markedly affect CH_4 emissions (Cai et al. 2003). In addition, N₂O is also emitted in large quantities from rice production systems following aerobic-anaerobic cycles; on the other hand, N₂O is emitted in short-term pulses after fertilisation, heavy rainfall or irrigation events and is the main GHG emissions of concern in upland wheat production systems (Wassmann et al. 2004). It is often difficult to balance emissions between CH_4 and N₂O when designing GHG mitigation strategies in a rice-wheat system, as measures to reduce CH_4 emissions often intensify N₂O emissions (Wassmann et al. 2004).

In evaluating the effect of intercropping on GHG emissions, Ricord (2018) studied GHG emissions from a sole maize crop, a sole soybean crop and a maize–soybean intercrop and found that the cereal–legume intercropping system effectively reduced N₂O emissions. In a similar study on the North China Plain, N₂O fluxes were lower from maize–soybean intercropping than a maize monoculture system in three growing seasons (2013–2015), when all cropping systems were applied with 240 kg N ha⁻¹ as urea in two split applications (Shen et al. 2018). Shen et al. (2018) showed that the fertiliser N loss as N₂O was lower in the maize-soybean intercropping (1.6%) and soybean monoculture (1.7%) than in the maize monoculture (2.3%), concluding that maize–soybean intercropping should be recommended as a climate-smart cropping systems for use on the North China Plain. A maize–wheat intercropping system coupled with reduced tillage and stubble mulching can increase grain production and

decrease C emissions in an arid area in northwest China (Hu et al. 2015; Yin et al. 2018). Therefore, choice of a cropping system to use and the associated management practices are important decisions to make to minimise GHG emissions.

8.3 Climate-Smart Agriculture (CSA) Practices and C Sequestration

Climate-smart agriculture has three key objectives: (1) to increase agricultural production per unit land area so as to increase income, food security and community development, (2) to improve the adaptive capacity at multiple levels, i.e. from the farm to the national level and (3) to reduce GHG emissions and to enhance C sinks in ecosystems (Campbell et al. 2014). Climate-smart agricultural management practices, including conservation tillage, use of cover crops, and biochar application to agricultural fields, has been widely considered a way to reduce GHG emissions from agriculture.

Conventional tillage has been identified as one of the causes of widespread land degradation problems, such as deterioration of soil structure, soil erosion and decreased soil fertility, affecting the long-term sustainability of agricultural production (Barber et al. 1996). Many climate-smart agricultural technologies have been tested to improve SOC storage in the agricultural landscape, and many of those have been demonstrated to be effective. Climate-smart agricultural technologies such as the use of cover crops, use of perennial crops, application of manure and biochar, reduced/minimum tillage or zero tillage, and crop rotation have all been shown to increase SOC storage.

Field experiments in Australia on light-textured soils in southern Australia indicate that conservation tillage (3–19 years in duration) was effective in increasing SOC levels as compared with conventional tillage, but only in areas with >500 mm annual precipitation and in the top 2.5-10.0 cm of the soil; the lack of conservation tillage effects on SOC levels in other climatic condition or soil layers was mainly attributed to low crop yield related to low rainfall, partial removal of stubble due to grazing and the high decomposition rate in areas with high air temperature (Chan et al. 2004).

8.4 Life Cycle Assessment (LCA) for Estimating the C Footprint of Agro-Food Systems

The proposal of effective GHG mitigation strategies in the agri-food sector needs to be based on a whole-system approach. This means that not only direct emissions but also indirect GHG losses (both upstream and downstream from the production

systems) must be considered. For that purpose, the calculation and use of the "C footprint" (CF) or "C budget" of agri-food entities, from products to systems, is primordial (highly important) (Plate 1.1).

The calculation of the CF of the agri-food system requires the accounting for GHG emissions that occur in each of the phases of food and feed production, including not only those that take place in the agricultural sector itself, that is, in crop fields and farms, but also during the manufacture of agricultural inputs, or those derived from the distribution, marketing and consumption of food, using an LCA approach.

The LCA adopts a "bottom-up" approach, that is, it records in detail the emissions generated along the product supply chain, using information on production technologies of the different goods and services. To this end, a "product system" has to be defined, which includes both the different phases of the supply chain (i.e. the "life cycle" of the product) and the exchanges that occur with the environment (i.e. GHG emissions); and a "functional unit" for each food (e.g. 1 kg of product). As a result of the application of an LCA, the "emission coefficients" (i.e. the amount of GHG emissions in kg CO_2 eq./kg of product) are obtained, which can be applied to both intermediate and final products, whether domestic or imported.

In estimating the CF of the Spanish Agri-food sector, Aguilera et al. (2015) used the following information as a source (Plate 8.3): (i) inventory analysis based on previous work of the Laboratory of History of Agroecosystems (UPO), based on official data (Yearbook of Agricultural Statistics, FAOSTAT, National Emissions Inventory, etc.); (ii) industrial input emission factors based on "embodied energy" (Aguilera et al. 2015); (iii) Mediterranean N₂O emission factors–meta-analysis (Cayuela et al. 2017); (iv) C sequestration with HSOC model (Aguilera et al. 2018); and (v) calculation of emissions associated with irrigation (Aguilera et al. 2019).

National inventories of atmospheric emissions, prepared by the signatory countries to verify compliance with the Kyoto Protocol, is the IPCC Tier 1 approach that is based on global emission factors. However, there is growing evidence that the



BOUNDARY OF THE PRODUCTION SYSTEM

Plate 8.3 An example of processes considered, and the main sources used to estimate the carbon footprint of the Spanish agri-food sector. Reproduced courtesy of Aguilera (2015)

factors differ depending on the type of climate, soil conditions and management, so it is necessary to use more specific factors for a more accurate assessment of emissions. The balance of C in the soil is a crucial process in the CF of agricultural products, since as we have seen it can compensate a large part of the emissions, when the soil gains organic matter and behaves as a sink; otherwise, the soil would contribute more to GHG emissions, when the soil loses organic matter. Despite this, it has been shown that, particularly under arid and semi-arid conditions, C in the soil is very sensitive to changes in management regimes, with changes that can range from the loss of half a ton of C per year in unfertilised soils, to the gain of more than one ton of C in soils in which organic amendments are applied.

The emissions derived from the production of agricultural inputs have decreased significantly due to improvements in efficiency in the industry; therefore, it is necessary to use temporal dynamic factors for the evaluation of historical GHG emissions. Quantitative reviews have been published that analyse precisely these factors for conditions comparable to those in European countries (Aguilera et al. 2015).

The sources of GHG emissions include the construction and maintenance of the agricultural infrastructure, the direct and indirect emissions associated with the use of energy, including traction animals, power generation and fuel use, and CH₄ emissions from water bodies (reservoirs, rafts, ditches and canals) (Aguilera et al. 2019). Emissions related to the use of energy could be estimated considering the changes in the country's electric mix, in the energy efficiency of electric generation, and in fossil fuel extraction techniques, including associated methane emissions. The GHG emissions associated with water bodies should include CH₄, CO₂ and N₂O.

Given the challenges that we face in reducing GHG emissions at the agri-food system level, it is necessary to advance our knowledge about effective mitigation strategies that are adapted to the soil-climatic conditions in each region, for example, by synthesising the existing relevant information regarding the main agricultural management practices and their impact on the mitigation of GHG emissions, C sequestration, other polluting compounds, as well as potential barriers and opportunities for the implementation of these strategies.

8.5 Conclusions

We conclude that CSA practices, with an emphasis on climate change adaptation and mitigation, can take many different forms. The CSA practices have many roles to play in agricultural sustainability and in reducing GHG emissions, as well as in increasing soil C sequestration. Practices such as the use of nitrification and urease inhibitors, mulching, application of biochar to the soil, fertilisation management and use of intercropping and crop rotations are all options available to landowners to effectively adapt to and mitigate regional to global climate change. The reader is, however, cautioned that the best CSA practice to be applied to a specific system or location depends on many different factors. Region- or site-specific research is often needed prior to their application to determine if any of the CSAs might produce a positive result on climate change adaptation and mitigation.

References

- Abalos D, Jeffery S, Sanz-Cobena A, Guardia G, Vallejo A (2014) Meta-analysis of the effect of urease and nitrification inhibitors on crop productivity and nitrogen use efficiency. Agric Ecosyst Environ 189:136–144
- Abalos D, Sanz-Cobena A, Misselbrook T, Vallejo A (2012) Effectiveness of urease inhibition on the abatement of ammonia, nitrous oxide and nitric oxide emissions in a non-irrigated Mediterranean barley field. Chemosphere 89:310–318
- Achtinich C, Bak F, Conrad R (1995) Competition for electron donors among nitrate reducers, ferric iron reducers, sulfate producers and methanogens in anoxic paddy soil. Biol Fertil Soils 19:65–72
- Aguilera E (2016) The influence of management practices on the greenhouse gas balance of Mediterranean cropping systems: identifying the climate change mitigation potential through quantitative review and life cycle assessment. PhD Thesis
- Aguilera E, Guzmán G, Alonso A (2015) Greenhouse gas emissions from conventional and organic cropping systems in Spain. II. Fruit tree orchards. Agron Sustain Dev 35:725–737
- Aguilera E, Guzmán GI, Álvaro-Fuentes J, Infante-Amate J, García-Ruiz R, Carranza-Gallego G, Soto D, González de Molina M (2018) A historical perspective on soil organic carbon in Mediterranean cropland (Spain, 1900–2008). Sci Total Environ 621:634–648
- Aguilera E, Vila-Traver J, Deemer BR, Infante-Amate J, Guzmán GI, González De Molina M (2019) Methane emissions from artificial waterbodies dominate the carbon footprint of irrigation: A study of transitions in the food-energy-water-climate nexus (Spain, 1900-2014). Environ Sci Tech 53:5091–5101
- Aulakh MS, Wassmann R, Rennenberg H (2001) Methane emissions from rice fields quantification, mechanisms, role of management, and mitigation options. Adv Agron 70:193–260
- Bai XX, Huang YW, Ren W, Coyne M, Jacinthe PA, Tao B, Hui DF, Yang J, Matocha C (2019) Responses of soil carbon sequestration to climate-smart agriculture practices: A meta-analysis. Glob Chang Biol 25:2591–2606
- Barber RG, Orellana M, Navarro F, Diaz O, Soruco MA (1996) Effects of conservation and conventional tillage systems after land clearing on soil properties and crop yield in Santa Cruz, Bolivia. Soil Tillage Res 38:133–152
- Barrena I, Menendez S, Correa-Galeote D, Vega-Mas I, Bedmar EJ, Gonzalez-Murua C, Estavillo JM (2017) Soil water content modulates the effect of the nitrification inhibitor 3,4-dimethylpyrazole phosphate (DMPP) on nitrifying and denitrifying bacteria. Geoderma 303:1–8
- Bodelier PLE, Roslev P, Henckel T, Frenzel P (2000) Stimulation by ammonium-based fertilizers of methane oxidation in soil around rice roots. Nature 403:421–424
- Bossio DA, Horwath WR, Mutters RG, van Kessel C (1999) Methane pool and flux dynamics in a rice field following straw incorporation. Soil Biol Biochem 31:1313–1322
- Bouwman AF (1990) Soils and the greenhouse effect. In: Proceedings of the international conference soils and the greenhouse effect. International Soil Reference and Information Centre (ISRIC), John Wiley and Sons, New York, pp 575
- Cai ZC, Mosier AR (2000) Effect of NH₄Cl addition on methane oxidation by paddy soils. Soil Biol Biochem 32:1537–1545

- Cai ZC, Sawamoto T, Li CS, Kang GD, Boonjawat J, Mosier A, Wassmann R, Tsuruta H (2003) Field validation of the DNDC model for greenhouse gas emissions in East Asian cropping systems. Glob Biogeochem Cyc 17:1107
- Cai ZJ, Gao SD, Xu MG, Hanson BD (2018) Evaluation of potassium thiosulfate as a nitrification inhibitor to reduce nitrous oxide emissions. Sci Total Environ 618:243–249
- Campbell BM, Thornton P, Zougmore R, van Asten P, Lipper L (2014) Sustainable intensification: What is its role in climate smart agriculture? Curr Opin Environ Sustain 8:39–43
- Cayuela ML, Aguilera E, Sanz-Cobena A, Adams DC, Abalos D, Barton L, Ryals R, Silver WL, Alfaro MA, Pappa VA et al (2017) Direct nitrous oxide emissions in Mediterranean climate cropping systems: emission factors based on a meta-analysis of available measurement data. Agric Ecosyst Environ 238:25–35
- Cayuela ML, Sanchez-Monedero MA, Roig A, Hanley K, Enders A, Lehmann J (2013) Biochar and denitrification in soils when, how much and why does biochar reduce N₂O emissions? Sci Rep 3:1732
- Chan KY, Heenan DP, So HB (2004) Sequestration of carbon and changes in soil quality under conservation tillage on light-textured soils in Australia: a review. Aust J Exp Agric 43:325–334
- Chang JY, Clay DE, Clay SA, Chintala R, Miller JM, Schumacher T (2016) Biochar reduced nitrous oxide and carbon dioxide emissions from soil with different water and temperature cycles. Agron J 108:2214–2221
- Conrad R, Rothfuss F (1991) Methane oxidation in the soil surface layer of a flooded rice field and the effect of ammonium. Biol Fertil Soils 12:28–32
- Daum D, Schenk MK (1998) Influence of nutrient solution pH on N₂O and N₂ emissions from a soilless culture system. Plant Soil 203:279–287
- Davidson EA, Ackerman IL (1993) Changes in soil carbon inventories following cultivation of previously untilled soils. Biogeochemistry 20:161–193
- FAO (2004) Carbon sequestration in dryland soils. World Soil Resources Reports 102. FAO, Rome, Italy, pp 108
- FAO (2013) Climate-smart agriculture sourcebook. Food and Agriculture Organization of the United Nations, Rome, Italy, p 570
- Feng Y, Xu Y, Yu Y, Xie Z, Lin X (2012) Mechanisms of biochar decreasing methane emission from Chinese paddy soils. Soil Biol Biochem 46:80–88
- Fentabil MM, Nichol CF, Neilsen GH, Hannam KD, Neilsen D, Forge TA, Jones MD (2016) Effect of micro-irrigation type, N-source and mulching on nitrous oxide emissions in a semi-arid climate: an assessment across two years in a Merlot grape vineyard. Agric Water Manag 171:49–62
- Flessa H, Wild U, Klemisch M, Pfadenhauer J (1998) Nitrous oxide and methane fluxes from organic soils under agriculture. Eur J Soil Sci 49:327–335
- Food and Agriculture Organization of the United Nations (2015) Soils: our ally against climate change [Video]
- Fuertes-Mendizábal T, Huérfano X, Vega-Mas I, Torralbo F, Menéndez S, Ippolito JA, Kammann C, Wrage-Mönnig N, Cayuela ML, Borchard N, Spokas K, Novak J, González-Moro MB, González-Murua C, Estavillo JM (2019) Biochar reduces the efficiency of nitrification inhibitor 3,4-dimethylpyrazole phosphate (DMPP) mitigating N₂O emissions. Sci Rep 9:2346
- Guo L, Wang X, Diao T, Ju X, Niu XG, Zheng L, Zhang X, Han X (2018) N₂O emission contributions by different pathways and associated microbial community dynamics in a typical calcareous vegetable soil. Environ Pollut 242(Pt B):2005–2013
- Holzapel-Pschorn A, Conrad R, Seiler W (1985) Production, oxidation and emission of methane in rice paddies. FEMS Microbiol Ecol 31:345–351
- Hu F, Chai Q, Yu A, Yin W, Cui HY, Gan YT (2015) Less carbon emissions of wheat-maize intercropping under reduced tillage in arid areas. Agron Sustain Dev 35:701–711
- Hüppi R, Neftel A, Lehmann MF, Krauss M, Six J, Leifeld J (2016) N use efficiencies and N₂O emissions in two contrasting, biochar amended soils under winter wheat-cover crop-sorghum rotation. Environ Res Lett 11:084013

- Hynšt J, Brůček P, Šimek M (2007) Nitrous oxide emissions from cattle-impacted pasture soil amended with nitrate and glucose. Biol Fertil Soils 43:853–859
- IPCC (2007) Summary for policymakers in Climate Change (2007) the physical science basis. In: Contribution of working group I to the fourth assessment report of the intergovernmental panel on climate change. Cambridge University Press, Cambridge, NY, USA, 2007
- IPCC (2014) Climate Change (2014) synthesis report. In: Core Writing Team, Pachauri RK, Meyer LA (eds) Contribution of working groups I, II and III to the fifth assessment report of the intergovernmental panel on climate change. IPCC, Geneva, Switzerland, pp 151
- IPNI (undated) Nitrification inhibitors. Nutrient source specifics(26)
- Ippolito JA, Laird DA, Busscher WJ (2012) Environmental benefits of biochar. J Environ Qual 41(4):967–972
- Karhu K, Mattila T, Bergström I, Regina K (2011) Biochar addition to agricultural soil increased CH₄ uptake and water holding capacity—results from a short-term pilot field study. Agric Ecosyst Environ 140(1–2):309–313
- Kniss AR, Savage SD, Jabbour R (2016) Commercial crop yields reveal strengths and weaknesses for organic agriculture in the United States. PLoS ONE 11(8)
- Knoblauch C, Maarifat A, Pfeiffer E, Haefele SM (2011) Degradability of black carbon and its impact on trace gas fluxes and carbon turnover in paddy soils. Soil Biol Biochem 43:1768–1778
- Kumar U, Jain MC, Pathak H, Kumar S, Majumdar D (2000) Nitrous oxide emission from different fertilizers and its mitigation by nitrification inhibitors in irrigated rice. Biol Fertil Soils 32:474–478
- Lam SK, Suter H, Bai M, Walker C, Davies R, Mosier AR, Chen DL (2018) Using urease and nitrification inhibitors to decrease ammonia and nitrous oxide emissions and improve productivity in a subtropical pasture. Sci Total Environ 644:1531–1535
- Lehmann J, Rillig MC, Thies J, Masiello CA, Hockaday WC, Crowley D (2011) Biochar effects on soil biota–a review. Soil Biol Biochem 43:1812–1836
- Li H, Liang XQ, Chen YX, Lian YF, Tian GM, Ni WZ (2008) Effect of nitrification inhibitor DMPP on nitrogen leaching, nitrifying organisms, and enzyme activities in a rice-oilseed rape cropping system. J Environ Sci 20:149–155
- Lindau CW, Bollich PK (1993) Methane emissions from Louisiana first and ratoon crop rice. Soil Sci 156:42–48
- Lipper L, Thornton P, Campbell BM, Baedeker T, Braimoh A, Bwalya M, Caron P, Cattaneo A, Garrity D, Henry K, Hottle R, Jackson L, Jarvis A, Kossam F, Mann W, McCarthy N, Meybeck A, Neufeldt H, Remington T, Sen PT, Sessa R, Shula R, Tibu A, Torquebiau EF (2014) Climate-smart agriculture for food security. Nat Clim Chang 4:1068–1072
- Liu Y, Yang M, Wu Y (2011) Reducing CH₄ and CO₂ emissions from waterlogged paddy soil with biochar. J Soils Sediments 11(6):930–939
- Lu WF, Chen W, Duan BW, Guo WM, Lu Y, Lantin RS, Wassmann R, Neue HU (2000) Methane emissions and mitigation options in irrigated rice fields in southeast China. Nutr Cycl Agroecosyst 58:65–73
- Ma J, Xu H, Yagi K, Cai Z (2008) Methane emission from paddy soils as affected by wheat straw returning mode. Plant Soil 313:167–174
- Majumdar D, Kumar S, Pathak H, Jain MC, Kumar U (2000) Reducing nitrous oxide emission from an irrigated rice field of North India with nitrification inhibitors. Agric Ecosyst Environ 81:163–169
- Matsushima M, Chang SX (2006) Vector analysis of understory competition, N fertilization, and litter layer removal effects on white spruce growth and nutrition in a 13-year-old plantation. For Ecol Manag 236:332–341
- Matsushima M, Chang SX (2007) Effects of understory removal, N fertilization, and litter layer removal on soil N cycling in a 13-year-old white spruce plantation infested with Canada bluejoint grass. Plant Soil 292:243–258

- Minasny B, Malone BP, McBratney AB, Angers DA, Arrouays D, Chambers A, Chaplot V, Chen Z-S, Cheng K, Das BS, Field DJ, Gimona A, Hedley CB, Hong SY, Mandal B, Marchant BP, Martin M, McConkey BG, Mulder VL, O'Rourke SM, Richer-de-Forges AC, Odeh I, Padarian J, Paustian K, Pan G, Poggio L, Savian I, Stolbovoy V, Stockmann U, Sulaeman Y, Tsui C-C, Vågen T-G, Van Wesemael B, Winowiecki L (2017) Soil carbon 4 per mille. Geoderma 292:59–86
- Mosier AR, Halvorson AD, Reule CA, Liu XJ (2006) Net global warming potential and greenhouse gas intensity in irrigated cropping systems in northeastern Colorado. J Environ Qual 35:1584– 1598
- Nägele W, Conrad R (1990) Influence of pH on the release of NO and N₂O from fertilized and unfertilized soil. Biol Fertil Soils 10:139–144
- Procházková B, Hrubý J, Dovrtěl J, Dostál O (2003) Effects of different organic amendment on winter wheat yields under long-term continuous cropping. Plant Soil Environ 49:433–438
- Ricord M (2018) Evaluating intercropping systems as a sustainable agroecosystem alternative to reduce greenhouse gas emissions. MSc thesis, University of Waterloo, pp 151
- Rodgers GA (1986) Nitrification inhibitors in agriculture. J Environ Sci Health A 21:701-722
- Rodriguez MJ, Saggar S, Berben P, Palmada T, Lopez-Villalobos N, Pal P (2019) Use of a urease inhibitor to mitigate ammonia emissions from urine patches. Environ Tech
- Rogers CW, Norman RJ, Brye KR, Slaton NA, Smartt AD (2015) Comparison of urease inhibitors for use in rice production on a silt-loam soil. Crop Forage Turfgrass Manag 1:1–6
- Sánchez-García M, Roig A, Sánchez-Monedero MA, Cayuela ML (2014) Biochar increases soil N₂O emissions produced by nitrification-mediated pathways. Front Environ Sci 2:25
- Sanderman J, Hengl T, Fiske GJ (2017) Soil carbon debt of 12,000 years of human land use. PNAS USA 114:9575–9580
- Sanz Cobena A, Misselbrook TH, Arce A, Mingot JI, Diez JA, Vallejo A (2008) An inhibitor of urease activity effectively reduces ammonia emissions from soil treated with urea under Mediterranean conditions. Agric Ecosyst Environ 126:243–249
- Sanz-Cobena A, Lassaletta L, Aguilera E, Del Prado A, Garnier J, Billen G, Iglesias A, Sánchez B, Guardia G, Abalos D, Plaza-Bonilla D, Puigdueta-Bartomoé I, Moral R, Galán E, Arriaga H, Merino P, Infante-Amate J, Meijide A, Pardo G, Álvaro-Fuentes J, Gilsanz C, Báez D, Doltra J, González-Ubierna S, Cayuela ML, Menéndez S, Díaz-Pinés E, Le-Noë J, Quemada M, Estellés F, Calvet S, van Grinsven HJM, Westhoek H, Sanz MJ, Gimeno BS, Vallejo A, Smith P (2017) Strategies for greenhouse gas emissions mitigation in Mediterranean agriculture: a review. Agric Ecosys Environ 238:5–24
- Sanz-Cobena A, Abalos D, Meijide A, Sanchez-Martin L, Vallejo A (2014) Soil moisture determines the effectiveness of two urease inhibitors to decrease N₂O emission. Mitig Adapt Strateg Glob Change
- Sanz-Cobena A, Sánchez-Martín L, García-Torres L, Vallejo A (2012) Gaseous emissions of N₂O and NO and NO₃leaching from urea applied with urease and nitrification inhibitors to a maize (Zeamays) crop. Agric Ecosyst Environ 149:64–73
- Sass RL, Fisher FM, Harcombe PA, Turner FT (1991) Methane emission from rice fields as influenced by solar radiation, temperature and straw incorporation. Glob Biogeochem Cyc 5:335–350
- Schütz H, Holzapfel-Pschorn A, Conrad R, Rennenberg H, Seiler W (1989) A three-year continuous record on the influence of daytime season and fertilizer treatment on methane emission rates from an Italian rice paddy field. J Geophys Res 94:16405–16416
- Schwartz J (2014) Soil as carbon storehouse: new weapon in climate fight? Yale School of Forestry & Environmental Studies
- Shen Y, Sui P, Huang J, Wang D, Whalen JK, Chen YQ (2018) Greenhouse gas emissions from soil under maize–soybean intercrop in the North China Plain. Nutr Cycl Agroecosyst 110:451–465
- Šimek M, Cooper JE (2002) The influence of soil pH on denitrification, progress towards the understanding of this interaction over the last 50 years. Eur J Soil Sci 53:345–354

- Šimek M, Jíšová L, Hopkins DW (2002) What is the so-called optimum pH for denitrification in soil? Soil Biol Bioch 34:1227–1234
- Skinner C, Gattinger A, Krauss M, Krause HM, Mayer J, van der Heijden MGA, Mäder P (2019) The impact of long-term organic farming on soil-derived greenhouse gas emissions. Sci Rep 9:1702
- Smith P (2016) Soil carbon sequestration and biochar as negative emission technologies. Glob Chang Biol 22:1315–1324
- Smith P, Martino D, Cai ZC et al (2008) Greenhouse gas mitigation in agriculture. Philos Trans R Soc B 363(1492):789–813
- Snyder CS, Bruulsema TW, Jensen TL, Fixen PE (2009) Review of greenhouse gas emissions from crop production systems and fertiliser management effects. Agric Ecosyst Environ 133:247–266
- Soares JR, Cantarella H, Menegale MLD (2012) Ammonia volatilization losses from surface-applied urea with urease and nitrification inhibitors. Soil Biol Biochem 52:82–89
- Souri MK, Neumann G (2018) Indications for passive rather than active release of natural nitrification inhibitors in *Brachiaria humidicola* root exudates. J Plant Nutrition 41:477–486
- Squalli J, Adamkiewicz G (2018) Organic farming and greenhouse gas emissions: a longitudinal US state-level study. J Clean Prod 192:30–42
- Stevens RJ, Laughlin RJ (1998) Measurement of nitrous oxide and di-nitrogen emissions from agricultural soils. Nutr Cycl Agroecosyst 52:131–139
- Subbarao GV, Nakahara K, Ishikawa T, Yoshihashi T, Ito O, Ono H, Ohnishi-Kameyama M, Yoshida M, Kawano N, Berry WL (2008) Free fatty acids from the pasture grass Brachiaria humidicola and one of their methyl esters as inhibitors of nitrification. Plant Soil 313:89–99
- Subbarao GV, Rao IM, Nakahara K, Sahrawat KL, Ando Y, Kawashima T (2013a) Potential for biological nitrification inhibition to reduce nitrification and N₂O emissions in pasture crop– livestock systems. Animal 7:322–332
- Subbarao GV, Nakahara K, Ishikawa T, Ono H, Yoshida M, Yoshihashi T, Zhu YY, Zakir HAKM, Deshpande SP, Hash CT, Sahrawat KL (2013b) Biological nitrification inhibition (BNI) activity in sorghum and its characterization. Plant Soil 366:243–259
- Subbarao GV, Rondon M, Ito O, Ishikawa T, Rao IM, Nakahara K, Lascano C, Berry WL (2007) Biological nitrification inhibition (BNI)—is it a widespread phenomenon? Plant Soil 294:5–18
- Tate KR, Ross DJ, Saggar S, Hedley CB, Dando J, Singh BK, Lambie SM (2007) Methane uptake in soils from Pinus radiata plantations, a reverting shrubland and adjacent pastures: effects of landuse change, and soil texture, water and mineral nitrogen. Soil Biol Biochem 39:1437–1449
- United Nations (2013) Greenhouse gas emissions and options for mitigation in Viet Nam, and the UNs responses
- Wang FH, Chen SM, Wang YY, Zhang YM, Hu CS, Liu BB (2018) Long-term nitrogen fertilization elevates the activity and abundance of nitrifying and denitrifying microbial communities in an upland soil: implications for nitrogen loss from intensive agricultural systems. Front Microbiol 9:2424
- Wassmann R, Lantin RS, Neue HU, Buendia LV, Corton TM, Lu Y (2000) Characterization of methane emissions from rice fields in Asia III: mitigation options and future research needs. Nutr Cycl Agroecosyst 58:23–36
- Wassmann R, Neue HU, Ladha JK, Aulakh MS (2004) Mitigating greenhouse gas emissions from rice-wheat cropping systems in Asia. In: Wassmann R, Vlek PLG (eds) Tropical agriculture in transition—opportunities for mitigating greenhouse gas emissions?. Springer, Dordrecht
- Waters D, Van Zwieten L, Singh BP, Downie A, Cowie AL, Lehmann J (2011) Biochar in soil for climate change mitigation and adaptation. In: Singh BP et al. (eds) Soil health and climate change. Soil biology, vol 29. Springer, Heidelberg, pp 345–368
- Wu FP, Jia ZK, Wang SG, Chang SX, Startsev A (2013) Contrasting effects of wheat straw and its biochar on greenhouse gas emissions and enzyme activities in a Chernozemic soil. Biol Fert Soils 49:555–565

- Xie Z, Xu Y, Liu G, Liu Q, Zhu J, Tu C, James SE, Amonette JE, Cadisch G, Jean WH, Yong JWH, Hu S (2013) Impact of biochar application on nitrogen nutrition of rice, greenhouse-gas emissions and soil organic carbon dynamics in two paddy soils of China. Plant Soil 370:527–540
- Yagi K, Tsuruta H, Minami K (1997) Possible options for mitigating methane emission from rice cultivation. Nutr Cycl Agroecosyst 49:213–220
- Yang M, Fang YT, Sun D, Shia YL (2016) Efficiency of two nitrification inhibitors (dicyandiamide and 3, 4-dimethypyrazole phosphate) on soil nitrogen transformations and plant productivity a meta-analysis. Sci Rep 6:22075
- Yin W, Guo Y, Hu FL, Fan ZL, Feng FX, Zhao C, Yu AZ, Chai Q (2018) Wheat-maize intercropping with reduced tillage and straw retention: a step towards enhancing economic and environmental benefits in arid areas. Front Plant Sci 9:1328
- Zaman M, Blennerhassett JD (2010) Effects of the different rates of urease and nitrification inhibitors on gaseous emissions of ammonia and nitrous oxide, nitrate leaching and pasture production from urine patches in an intensive grazed pasture system. Agric Ecosyst Environ 136:236–246
- Zaman M, Nguyen ML (2010) Effect of lime or zeolite on N₂O and N₂ emissions from a pastoral soil treated with urine or nitrate-N fertiliser under field conditions. Agric Ecosyst Environ 136:254–261
- Zaman M, Nguyen ML, Saggar S (2008a) N_2O and N_2 emissions from pasture and wetland soils with and without amendments of nitrate, lime and zeolite under laboratory condition. Aust J Soil Res 46:526–534
- Zaman M, Nguyen ML, Gold AJ, Groffman PM, Kellogg DQ, Wilcock RJ (2008b) Nitrous oxide generation, denitrification and nitrate removal in a seepage wetland intercepting surface and subsurface flows from a grazed dairy catchment. Aust J Soil Res 46:565–577
- Zaman M, Nguyen ML, Matheson F, Blennerhassett JD, Quin BF (2007) Can soil amendments zeolite or lime shift the balance between nitrous oxide and dinitrogen emissions from pasture and wetland soils receiving urine or urea-N? Aust J Soil Res 45:543–553
- Zaman M, Saggar S, Blennerhassett JD, Singh J (2009) Effect of urease and nitrification inhibitors on N transformation, gaseous emissions of ammonia and nitrous oxide, pasture yield and N uptake in grazed pasture system. Soil Biol Biochem 41:1270–1280
- Zhu GD, Ju XT, Zhang JB, Muller C, Rees RM, Thorman RE, Sylvester-Bradley R (2019) Effects of the nitrification inhibitor DMPP (3,4-dimethylpyrazole phosphate) on gross N transformation rates and N_2O emissions. Biol Fertil Soils 55:603–615
- Zvomuya F, Rosen CJ, Russelle MP, Gupta SC (2003) Nitrate leaching and nitrogen recovery following application of polyolefin coated urea to potato. J Environ Qual 32:480–489

The opinions expressed in this chapter are those of the author(s) and do not necessarily reflect the views of the International Atomic Energy Agency, its Board of Directors, or the countries they represent.

Open Access This chapter is licensed under the terms of the Creative Commons Attribution 3.0 IGO license (http://creativecommons.org/licenses/by/3.0/igo/), which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the International Atomic Energy Agency, provide a link to the Creative Commons license and indicate if changes were made.

Any dispute related to the use of the works of the International Atomic Energy Agency that cannot be settled amicably shall be submitted to arbitration pursuant to the UNCITRAL rules. The use of the International Atomic Energy Agency's name for any purpose other than for attribution, and the use of the International Atomic Energy Agency's logo, shall be subject to a separate written license agreement between the International Atomic Energy Agency and the user and is not authorized as part of this CC-IGO license. Note that the link provided above includes additional terms and conditions of the license.

The images or other third party material in this chapter are included in the chapter's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the chapter's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder.

