

Review

Toxicological Issues of Nanoparticles Employed in Photocatalysis

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Abstract. A huge amount of different nanomaterials is nowadays on the market used for various specific applications. Some nanomaterials such as TiO₂, ZnO as well as several other semiconductors exhibit photocatalytic activity.

Hence these materials are used for many applications, e.g., for self-cleaning and antibacterial coatings on different surfaces and for the purification of wastewater where the cleaning can be induced by simple exposure to sunlight. Because of the frequent use of these nanoparticles it is important to investigate the life cycles of these nanostructured materials as well as their environmental impact and their toxicity to animals and humans.

This review first gives a short overview about nanotechnology and nanotechnological products as well as about photocatalysis and semiconductors used in this field. We then discuss the need for a new technology named nanotoxicology and the problems occurring when investigating the toxic potential of nanomaterials as well as the life cycle of nanomaterials. Furthermore, we focus on the environmental impact of TiO₂ and ZnO nanoparticles including toxic effects to bacteria, water organisms and plants as well as their toxic effects to humans including in vitro and in vivo studies.

Keywords. Nanotoxicology, nanotechnology, TiO₂ ZnO, photocatalysis, environmental impacts, human health risks, in vivo studies, in vitro studies.

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1 Nanotechnology

The technological progress allows the production of new tiny materials which have the size of only a few nanometers. Nanotechnology deals with the production, investigation and utilization of these extremely small particles [1].

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Nanotechnology is projected to become one of the key technologies of the 21st century [2]. In the nanometer range several physical and chemical properties of materials change which can be specifically used for many applications.

Nanoparticles have an extremely high surface to volume ratio and new effects, properties and functions can be observed in the nanometer scale [3]. Mechanical, optical, magnetic, electrical and chemical properties are not only depending on the material but also on the size and shape of the particles. New phenomena occur which cannot be explained by the laws of classical physics, e.g., quantum effects start playing a role as the particles get very small.

The fabrication of nanomaterials can be achieved employing the top-down or the bottom-up method, respectively. With the top-down process nanoscale materials are produced by miniaturization of any type of source material. The nanoparticles can be build up to more complex systems or can be mixed to other materials to modify their properties. Today this approach dominates in physics and physical techniques with the modern semiconductor industry being just one example. However, the constant development of miniaturization in the semiconductor industry is limited and is not able to follow the increase of the number of transistors [4]. Moreover, the top-down approach is an energy extensive, waste producing and resource wasting process.

Using the bottom-up approach nanomaterials are assembled from atoms or molecules. This approach is frequently used in chemistry and biology. The nanoparticles and their assemblies are, for example, synthesized by chemical vapor deposition or by crystal growth from suitable seeds. One promising approach is the growth by self-assembly [5], [6]. Typical materials synthesized employing the bottom-up process are carbon nanotubes and nanocrystalline zeolites. Since the bottom-up and the self-assembly process can be used in almost every scale these methods are extremely powerful.

Potential applications of nanomaterials can be found in nearly all sectors of industry. In the information and communication sector nanotechnology is specifically important for the production of processors because of the resulting miniaturization. Furthermore, photonic crystals with their potential application in the area of optical circuitry are of great interest for information processing based only on light (photonics).

Nanostructured materials can also be adopted for medical and health care applications [7], e.g., as biocompatible materials produced for the use as implants and for tissue engineering [8], [9]. Moreover, drugs can be transported in the organism and released on target (drug delivery) [10],

Industry Sector	Application
Chemistry	Paints [21], Catalysts [22], [23], Hydrogen Storage [24]
Automotive Industry	Wheel Fillers, Antireflection Coatings [25], Scratch-Resistant Paints, De-Mister Coatings, Fuel Cells
Optical Industry	White LEDs, Quantum Dot Laser, Photonic Crystals,
Medicine and Health Care	Implant Materials [8], [9] <i>Drug Delivery</i> , [10], [11] <i>Functional Foods</i> [26], Cosmetics, Fluorescent Biological Labels [12], [13], [14], Detection of Pathogens [15] Proteins [16], Hyperthermia [18], Antibactericidal Surfaces [21]
Environmental Technology	Water Treatment [27], Solar Energy [28], Conversion and Storage Systems, Pollution Control and Abatement
Information and Communication Technology	Processors

Table 1. Applications of nanoparticles.

[11], and the availability of bioactive substances can be increased (functional foods). Nanoparticles can also be used as fluorescent labels [12], [13], [14], and for the biodegradation of pathogens [15] and proteins [16]. Another application of nanotechnology is the separation and purification of biological molecules and cells [17]. A promising and important use in medicine is the selective tumor destruction with nanomaterials via heating called hyperthermia [18].

In the automotive industry nanotechnological products are used as wheel fillers, antireflection coatings, scratch-resistant paints and hydrophilic coatings as de-mister surfaces. Nanoscale materials are used in photovoltaic devices and fuel cell for power supplies. The application of nanotubes in lithium-ion-accumulators can increase the capacity of batteries and mini-accumulators. In the construction industry the application of nanoscale materials in metals enables an essential contribution to light construction. The admixing of silica nanoparticles to construction materials enhances some material properties such as adhesive pull strength and adhesive shear strength between concrete and armoring steel. In environmental technology nanoparticles are used as catalysts for water treatment and for the elimination of pollutants. The underlying mechanism and the employed materials for this purpose will be discussed in more detail in the following section (photocatalysis). It is obvious that nanotechnology products have a wide range of different applications and have already entered their commercial exploration period [19], [20].

Nanostructured materials already exist much longer than the rather recent emergence of nanotechnology. Many long-existing materials are in fact structured on the micro- and nanometer scale and many industrial processes that have been used for decades such as polymer and steel manufacturing exploit nanoscale phenomena. Natural processes have produced nanoparticles for eons for example volcanic eruptions. Nanometer-sized particles are created in countless physical processes like erosion and combustion

and the natural world is full of examples of systems with nanoscale structures, e.g., proteins, cells, bacteria, viruses, etc.

Because nanomaterials are used in many different applications and are generated as by-products in technical processes it is important to investigate their potential environmental risks as well as the possible hazards to animals and human beings.

2 Photocatalysis and Photocatalysts

The fundamental principle of semiconductor photocatalysis is the ability of the employed catalyst particles to absorb photons to create reactive electron-hole-pairs which are capable of oxidizing most organic and inorganic compounds. This phenomenon is the basis for a variety of current and projected applications in different fields, such as surface technology, pollution management and medicine.

Briefly, when a semiconducting material absorbs a photon with an energy equal or higher than its bandgap energy, an electron is excited from the valence to the conduction band. Simultaneously, a positively charged hole (h^+) is created in the valence band [29]. Recombination of these two charge carriers will result in the generation of heat or in the reemission of a photon. However, once these two charge carriers reach the surface of the semiconductor, they can undergo a variety of reactions with surface-adsorbed molecules (see Figure 1). For many applications, the most relevant reaction is the oxidation reaction, as the photogenerated hole in the valence band of, i.e., the very frequently used photocatalyst titanium dioxide (TiO_2), has a sufficiently high redox potential to be able to oxidize most organic compounds. In addition to the direct reaction with the holes, this oxidation reaction can be mediated by several active oxygen species. These mediators include but are not limited to hydroxyl radicals ($\bullet OH$), superoxide radicals ($O_2^{\bullet -}$), and singlet oxygen (1O_2) [30], [31], [23], [32], [33].

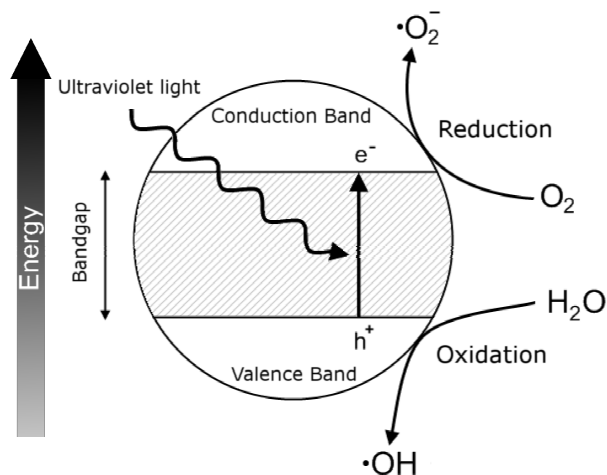


Figure 1. Illustration of the fundamental principle of photocatalysis.

Similarly, the conduction band electron can react with adsorbed molecular oxygen forming superoxide radicals and subsequently hydrogen peroxide (H_2O_2). However, the oxidizing potential of photocatalytic materials is not limited to their surface, as oxidizing species can diffuse away from the photocatalytic surface, inducing redox reactions in the bulk. This effect is usually called remote photocatalysis and is most likely mediated through hydrogen peroxide and singlet oxygen [34], [35], [36], [37], [38], [39], [40], [41], [42], [43], [44], [45], [46], [27]. However, in the case of remote photocatalysis, the reaction rate and thus the product yield is usually one or two orders of magnitude lower than that of the process occurring at the photocatalyst surface. In many cases, only the initial oxidation reaction is photocatalytic, forming a reactive radical that is further oxidized by molecular oxygen, eventually resulting in the formation of carbon dioxide [47]. Besides this exergonic reaction sequence, photocatalytic fuel synthesis, e.g., the cyclic splitting of water into molecular hydrogen and oxygen, presents another very interesting application of photocatalysis that currently receives increased attention and has been the topic of several recently published reviews [48], [49], [50].

Given that photocatalysis is a surface phenomenon, a high surface area potentially contributes to a high photocatalytic activity. Additionally, if the particles are small enough, their light scattering properties become negligible and suspensions or coatings made from these particles will appear optically transparent, which is an important property for many applications [51]. For these reasons, preferably materials exhibiting a high specific surface area and minute dimensions are used in applications, specifically nanoparticles and nanostructured materials [52].

Although sometimes non-oxidic compounds (CdSe, ZnS, GaP) are used as photocatalysts, their instability in aqueous environments and their susceptibility to photo-

corrosion greatly limits their applications [53], [27], [54]. Therefore, most applications employ the more stable metal oxide semiconductors such as TiO_2 , ZnO, and in some cases, Fe_2O_3 and WO_3 as photocatalysts. Unfortunately, due to their rather high bandgap energy of 3.2 eV, the use of TiO_2 and ZnO is limited to light in the ultraviolet region, i.e., below 400 nm. Ultraviolet light, however, only constitutes a small portion of the sunlight. For this reason, the catalysts are sometimes modified by doping, e.g., incorporating N, C, and or S atoms into the bulk material or by surface modification, e.g., depositing noble metal islands as electron transfer catalysts in order to achieve better photocatalytic activity and/or an activity under visible light irradiation.

So far photocatalysis has been proposed for or applied in a variety of applications. Currently, the two most important application fields are surface technology and waste management. In surface technology, surfaces are coated with photocatalytic material to achieve several different properties. One of these properties is the so-called self-cleaning effect: organic pollutants are photocatalytically degraded on surfaces coated with photocatalysts as long as they are illuminated with a sufficient amount of ultraviolet light. This technique is applied to glass surfaces, outdoor building façades, roof tiles, ceramics, plastics and even fabrics and wool [55], [56], [57], [25], [58]. Since the surface can not only be cleaned from organic pollutants but can also be disinfected, the photocatalytic technology can also be used for medical instruments or for surfaces in medical facilities [59], [60], [61].

Another important application of photocatalytic surfaces is the decontamination, deodorization and disinfection of indoor air. Interior facilities are often contaminated with malodorous compounds that reduce the living quality of the residents. Additionally, construction materials and furniture often emit volatile organic compounds (VOCs) in low concentrations. These VOCs may be harmful for the inhabitants and cause diseases such as the *sick building syndrome*. Since almost all airborne pollutants can be degraded by photocatalysis, using this technique to clean the air is more reliable and needs less maintenance than conventional filter-based air-purification systems. Furthermore, since bacteria, fungal spores and viruses are also degraded in this process, they can help to reduce the number of infections transmitted in hospitals and other medical facilities.

Likewise, this method can also be applied for outdoor air purification. The main aim here is, however, to reduce environmental pollutants such as nitrous oxides. Construction materials, roads and pavements are thus modified with photocatalytic material in an attempt to reduce pollutant levels in high-traffic regions [62], [63], [64], [65], [66].

Photocatalysis can also play an important role in industrial and communal waste water management. The advantage of using photocatalysis for waste water treatment is that, unlike in other advanced oxidation techniques, besides

air no external reagent (e.g., O₃, H₂O₂) is needed for the reaction. This makes photocatalysis a potentially cheaper and easier to operate alternative to conventional advanced oxidation techniques for the decontamination of waste water [27], [67], [68], [69], [70], [71]. However, besides the engineering of appropriate reactors the main current drawback for a wider application of this technology for water treatment is the lack of photocatalysts exhibiting sufficient and long-lasting activity and/or being able to utilize also visible light.

3 Nanotoxicology

Currently, a wide range of nanoparticles of different types and exhibiting different properties are being synthesized which are going to be tested for their suitability for various industrial applications. As more and more technological products are becoming available to the customers, concerns have been raised regarding the harmlessness and the potential toxicity of these materials [72], [73], [74]. Every technology or designed product requires careful investigations regarding its sustainability and the associated risk potential before being introduced into the market. For this reason, a new subdiscipline of nanotechnology namely nanotoxicology emerged [75], [76]. Nanotoxicology was initially defined as the science dealing with the effect of engineered nanomaterials and nanostructures on living organisms emerging from the toxicology of ultrafine airborne particles [75]. If the definition of toxicology is adapted to nanomaterials, nanotoxicology can be described as the science dealing with the environmental impact and the hazard of nanomaterials for living organisms as well as the interaction of these materials with biological and environmental systems, including the prevention and minimization of adverse effects caused by nanomaterials [74].

Various new difficulties and subsequent new approaches result from the investigation of the toxicity of nanostructured materials, because compared to their bulk counterparts nanomaterials exhibit a different behavior resulting from property changes in the nanoscale regime.

One of the important factors in nanotoxicology is the possible exposure to nanoparticles and the life cycle of nanostructured materials. The main problem in the determination of the exposure is the lack of labeling of products containing nanomaterials and the lack of knowledge of the concentration of these materials in the environment. Appropriate measurements of particle concentrations and doses and the knowledge of produced materials and their disposal in the environment therefore constitute the first prerequisite for this new discipline. Natural airborne particles in the environment add to the extremely complicated task of the determination of anthropogenic nanoparticles. There are some approaches to determine, to calculate and, or to measure the number of particles in a given environment [77], [78], [79],

[80], [81]. Furthermore, since workers and consumers have to be protected, a labeling of nanomaterials will be needed.

Toxicity testings of nanomaterials in vivo or in vitro by establishing dose-response relationships have been developed to identify a potential hazard. Because risk is a function of hazard and exposure [risk = f (hazard, exposure)], the approach is to incorporate both components into a paradigm. Therefore, hazard identification has to be achieved and consequently dose-response and exposure assessment must be carried out. Following these steps the risk assessment and finally the risk management can be executed [83], [84].

The characterization of nanomaterials is very important and several nanoparticle classes have to be identified. The physiochemical characteristics and the biological reactivity of particles play important roles. The chemical structure alone is not sufficient to characterize nanomaterials, but rather shape, size (ferret/hydrodynamic), size distribution, agglomeration/aggregation, stability, solubility in different media, surface properties (area/porosity, charge, reactivity, coatings/contaminants, defects), composition and crystal structure have to be determined to enable a judgment concerning the possible hazards of these materials. These properties can change with the method of production, the preparation process, the storage of materials and the introduction into physiological media and the organism. It is not possible to compare the toxicity of nanomaterials to that of their bulk counterparts, because as described in Section 1 the reactivity and the material properties are not identical. Size is an important parameter to predict entry routes, efficiency and translocation of nanoparticles in the organism. Their surface charge may contribute to possible molecular interactions.

Due to their small size, the particles can usually be easily incorporated into the human body. The most important and most investigated adsorption pathway is the assimilation via the lung. Several studies highlight the possible hazard of lung diseases caused by nanomaterials reaching the respiratory tract [85], [86], [87], [88]. Exposition and inhalation studies show the incorporation of particles in the lung and the localization and possible elimination of particles depending on their size, shape, aggregation and surface area [89], [90], [91], [92]. A considerable amount of research demonstrates the toxicity of combustion-derived particles such as diesel soot [90], [91], welding fume [92], carbon black [93], and coal fly-ash [94]. However, there are also studies concerning the incorporation of nanoparticles via the skin [95]. Until now, there is no evidence that the nanoparticles can enter the organism via skin adsorption. Usually the particles are located in the horny layers while only a very small fraction of nanoparticles such as TiO₂ and ZnO used in sunscreens was found to penetrate into deeper tissue layers but only inside single follicle channels. [96], [97], [98], [99]. Luke et al., however, showed that Quantum Dots are able to penetrate the skin [100].

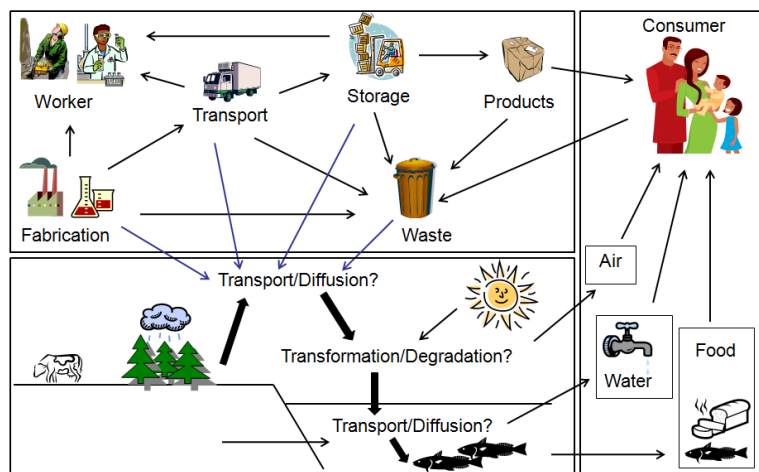


Figure 2. : Scheme of the life cycle of nanomaterials [82].

Besides these “natural” or accidental adsorption pathways of nanoparticles into the organism, intended insertion of nanoparticles into the body occurs for medical applications either intravenously [101] or when present as a layer on bone tissue implants [8], [9].

After entering the organism, nanoparticles will most likely be able to move freely because of their small size thus reaching almost every part of the body. Various studies have shown that the localization of particles in the body depends on their adsorption path in the organism [102] as well as on their size and shape [103]. The effective clearance mechanism of particles that had entered the tissues was also found to depend on their size [104], [105]. Translocation and toxicity are also regulated by specific cellular interactions and by specific nanomaterial factors, such as their biochemical stability and their potential to generate reactive oxygen species (ROS) [106]. ROS can cause oxidative stress which has been linked to increased cytotoxicity [107], [108], [109], [110].

Nanoparticles can interact with various biological molecules such as proteins [111], [112] and are able to penetrate the cell membrane interacting with DNA and causing DNA damage [113]. Evidence has also been presented that nanoparticles can break through the blood-brain-barrier [95], [114]. The surface charge will influence nonspecific adhesion of proteins on the nanomaterials surface as well as its permeation through junctions and its cellular uptake [104].

As mentioned before, particles can react with proteins. Therefore, the particle size, the aggregation and the agglomeration state in the biological media used for cell culture tests has to be determined. It is important to verify that during any test nanoparticles and not only aggregates of particles are still in the culture media [115].

Several studies show that particles exhibit a different behavior in the culture medium as compared to aqueous sus-

pensions. Some particles are stabilized by the proteins in the cell culture media while some are aggregated by salts [116], [117], [118], [119], [120].

This overview concerning nanotoxicology research illustrates the difficulties in applying generalized concepts for the prediction of nanomaterial permeation, of cellular uptake and of toxicity in biological systems. Nanotoxicology is an emerging field with a lot of antagonisms existing in the published literature since standardized tests do currently not exist.

4 Environmental Impacts of Nanoparticles

The environmental impact caused by nanomaterials is presently being extensively discussed in particular in correlation with the regulations concerning airborne particulate matter. To be able to estimate the potential risk of nanomaterials informations concerning their mobility and the life cycle are of crucial importance.

Furthermore, a clear differentiation between particles generated by technical processes and those unintended released is definitely required. Most of the latter particles are generated by combustion processes (flow ash, diesel soot, catalyst dust, carbon black).

Naturally occurring nanoparticles vary in size, composition and shape, whereas artificial nanomaterials are more uniform because they are synthesized for special applications. With the progressive development of traffic a drastic increase of ultrafine particles in the air is noticeable. The life cycle of nanomaterials is not obvious, cf. Section 3. Depending on production and use, nanomaterials can be emitted into air or water and can thus be found on the ground or in ground or surface water, respectively.

Investigations of combustion-derived ultrafine particles can assess the behavior and the impact of nanoparticles

in the environment. The ultrafine particles produced by diesel engines have the property to agglomerate and to subsequently sediment within a few days, hence, these dusts are eliminated from the air after a certain period of time. Synthesized, artificial particles frequently do not agglomerate because they are often stabilized with, e.g., core-shell-systems for specific applications; consequently, they stay in the air for a longer period of time and can travel over long distances.

Because of their large surface area nanoparticles can adsorb more pollutants and bigger amounts of toxic substances such as pesticides, fertilizer, pollutants and heavy metals, which can thus be transported over long distances and can be widely distributed [121]. Due to their high reactivity nanoparticles may react with natural substances possibly forming toxic compounds or composites. Furthermore, the bactericidal effect of some materials could change the microbial composition in the water or on the ground. To assess the environmental hazard, most studies investigate the impact and toxic effects of nanomaterials to organisms living in water. The first step in an investigation of this exposure route is the characterization of the form and the concentration of engineered nanomaterials in water since this is where many substances exhibit their most significant environmental impact. Materials present in water can be degraded, transformed and/or accumulated in a variety of ways.

In the following we will focus on the environmental effects caused by semiconductors such as TiO_2 and ZnO used in photocatalysis. The life cycle of these materials is long. They can be released to the environment from laboratories and factories during their production processes. Because of the wide use as photocatalysts release to the environment is also possible from products containing these materials, such as photocatalytic wall paints and coated surfaces, e.g., roof tiles. In some products the photocatalyst particles are present in solutions, e.g., during the treatment of wastewater. In others they are present as fixed particles in coatings. TiO_2 and ZnO nanoparticles are being prepared in large quantities and in many different varieties; some are even stabilized with proteins and other reagents. As mentioned before, the bactericidal effect of materials can also have an environmental impact.

Bactericidal Effect of TiO_2 and ZnO Nanoparticles

The toxic effect of illuminated TiO_2 and ZnO nanoparticles is being thoroughly investigated and many papers have been published on this topic with the majority of these studies being performed with TiO_2 because this is by far the most used photocatalyst.

When exposed to near-UV light, titanium dioxide exhibits a strong bactericidal activity. However, the underlying killing mechanism of this photocatalytic reaction is not yet well understood. Recently, total oxidation of *Esche-*

richia coli cells has been demonstrated [122]. The reactive oxygen species (ROS) generated by the TiO_2 photocatalytic reactions cause various damages to living organisms. This is not surprising since the latter are composed of organic compounds. In 1985, Matsunaga and coworkers reported the microbiocidal effect of TiO_2 photocatalytic reactions for the first time [123].

E. coli has been chosen in many studies to investigate the toxic effect of photocatalysis [124], [125], [126]. Coleman et al. examined the photocatalytic degradation of *E. coli* in water comparing various catalysts and showed that TiO_2 (P25) was the most efficient catalyst. Non-buffered water samples displayed a greater bactericidal efficiency, which was attributed to a decrease in the electrostatic repulsion between TiO_2 and *E. coli* and to elevated stress on *E. coli* at acidic pH. Buffered samples showed a decrease in bactericidal efficiency which was attributed to the presence of HCO_3^- -ions competing for oxidizing species and blocking the surface of the TiO_2 -particles [124].

It has also been shown that TiO_2 photocatalyst particles illuminated with UV-light exhibit high disinfection rates against other microorganisms besides *E. coli* such as *Lactobacillus acidophilus* and *Saccharomyces cerevisiae* [123], *Lactobacillus helveticus* [127], *Pseudomonas aeruginosa* [128] *Pseudomonas stutzeri* [129] and *Bacillus pumilus* [130].

The toxic effect of photocatalytic systems to bacteria such as *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Enterococcus faecium* and *Candida albicans* which are relevant for hygiene has also been investigated [131]. Kuhn et al. observed a high mortality for all these bacteria on UV-illuminated surfaces coated with TiO_2 (P25) and proposed that $\bullet\text{OH}$ -radicals formed on the catalyst surface are causing this effects [131].

Gogniat et al. analyzed the bactericidal effect of UV-illuminated TiO_2 in NaCl-KCl or sodium phosphate solutions to an *E. coli* strain. They found that the adsorption of bacteria on the catalyst occurred immediately in NaCl-KCl solution, whereas it was delayed in the sodium phosphate solution. Moreover, the adsorption rate of the cells onto the catalyst was found to be positively correlated with its bactericidal effect. The authors suggest that the photocatalytic bactericidal effect is related to the loss of membrane integrity of the cells aggregated on the TiO_2 surface [132].

The toxic effect of TiO_2 photocatalysis to phytopathogenic bacteria is not only a chance to alternatively control plant diseases rather than using pesticides, but it can also present a hazard which currently cannot yet be estimated [133], [134].

The photocatalytic activity of ZnO nanoparticles on different microorganisms has also been reported [135], [136], [137], [138]. Upon UV(A)-illumination ZnO causes a significant growth inhibition to *E. coli* [136], [137], [138], *Bacillus subtilis* [139], *Streptococcus agalactiae*, and *Staphylococcus aureus*, with the latter two being patho-

genetic agents causing several infective diseases [135]. The antibactericidal activity of ZnO to *Staphylococcus epidermidis*, *Streptococcus pyogenes*, *Enterococcus faecalis* [139] has also been investigated. Jones et al. found the antibacterial activity of ZnO nanoparticles to depend on their size, with smaller particles exhibiting a greater efficacy for the bacteria growth inhibition.

While in the absence of UV illumination TiO₂ is not able to inhibit the growth of, e.g., *Staphylococcus aureus* significantly, ZnO particles are found to also exhibit bactericidal activity in the dark [140], [141].

Brayner et al. observed that synthesized ZnO nanoparticles with average particle diameters of 12 nm are able to inhibit the bacterial growth (100% at concentrations of 3 mM) and suggested this to be due to the disorganization of the *E. coli* membranes. As a consequence the membrane permeability increases leading to the accumulation of nanoparticles in the bacterial membrane and the cytoplasm of the cells [142].

Roselli et al. proposed that the toxic effect of ZnO to *E. coli* is related to zinc ions which are metabolized as an oligo-element [138]. Certain bacteria have developed mechanisms to regulate the influx and efflux processes to maintain a steady intracellular concentration of metal ions, including the Zn²⁺ ion. The genes responsible for the transport of zinc ions have also been characterized in several bacteria, including *Streptococcus pneumoniae*, *Haemophilus influenzae*, *Staphylococcus aureus*, *E. coli*, and *Bacillus subtilis* [143], [144].

Impacts of TiO₂ and ZnO Nanoparticles on Water Organisms and Plants

The effect of nanoparticles on water organisms and plants also plays an important role concerning the environmental impact of nanotechnological products, because nanoparticles may be released into freshwater systems as a result of their use for the treatment of waste from industry and medicine. There are only a few studies investigating these interactions. Free radicals generated by the irradiation of photocatalysts may cause a risk for organisms living in water.

Hundt-Rinke et al. investigated the effect of two different nanoparticle suspensions under appropriate illumination to induce their photocatalytic activity. The growth inhibition of the green alga *Desmodesmus subspicatus* and the immobilization of *Daphnia magna* were selected as test reaction, respectively. It was shown that nanoparticles may induce ecotoxicological effects depending upon the nature of these particles. TiO₂ particles with a size of 25 nm mainly consisting of anatase exhibited a harmful effect on algae (EC₅₀-values 44 mg/L), whereas pure anatase with a size of 100 nm show no toxic effect to the algae (maximum investigated concentration 50 mg/L). In the tests performed with daphnia, toxicity was observed for both TiO₂ nanoparticles

[145]. Lovorn and coworkers also investigated the toxic effect of TiO₂ to daphnia and reported an increase in mortality with an increase of the nanoparticle concentration [146].

Velzeboer et al. investigated the ecotoxic effect of some nanomaterials employing different methods and ecotoxicity endpoints. Four different ecotoxicity tests were performed: the Microtox[®] test, the pulse-amplitude modulation (PAM) test, the Chydotox test, and the Biolog[®] test [147]. The Microtox[®] test uses bioluminescent bacteria, specifically, the strain *Vibrio fischeri* [148], the pulse-amplitude modulation (PAM) uses the green alga *Pseudokirchneriella subcapitata* [149], the Chydotox is based on the survival of *Chydorus sphaericus*, a small, benthic cladoceran [150], and the Biolog[®] test uses a mix of soil bacteria to determine toxicity via multivariate analysis [151]. No appreciable effects were observed at concentrations up to 100 mg/L for tested particles (TiO₂, ZrO₂, Al₂O₃, CeO₂, fullerene (C₆₀), single-walled carbon nanotubes, and polymethylmethacrylate). It was suggested that colloid (in)stability is of primary importance in explaining ecotoxic effects of nanoparticles in the natural environment [147].

The toxicity of TiO₂ nanoparticles to the alga *Pseudokirchneriella subcapitata* was also investigated with bulk TiO₂ (EC₅₀ = 35.9 mg/L / NOAEC = 10,1 mg/L) showing a lower toxicity as compared to their nano formulations (EC₅₀ = 5.83 mg/L / NOAEC = 0,98 mg/L). Nano TiO₂ formed characteristic aggregates entrapping algal cells which may contribute to the toxic effect of nanostructured TiO₂ to algae [152].

On the other hand, suspensions of nano and bulk TiO₂ were found to be non toxic even at loadings at 20 g/L to bacteria *Vibrio fischeri* and to crustaceans *Daphnia magna*, and to *Thamnocephalus platyurus* [153]. This study did not include any measurements of the TiO₂ particle size in water. Commercially available TiO₂ nanoparticles e.g., Evonik-Degussa Aeroxide P25 showed no measurable effect on the growth rates of four different phytoplankton species: *Thalassiosira pseudonana*, *Skeletonema marinoi*, *Dunaliella tertiolecta*, and *Isochrysis galbana*, up to concentrations of 1000 µg/L (ppb) [154].

Zinc oxides were found to be equally toxic to the alga *Pseudokirchneriella subcapitata* in their bulk and in their nano formulations [151]. The toxicity of ZnO was attributed to soluble zinc ions originating from the metal oxide particles. This study by Aruoja et al. again demonstrates that solubility appears to be a key issue governing the toxicity of metal containing nanoparticles, at least for organisms that a priori are not internalizing the particles [152].

Heinlaan and coworkers investigated the toxic effect of nano and bulk ZnO as well as of Zn²⁺ ions to bacteria *Vibrio fischeri*, to crustaceans *Daphnia magna* and *Thamnocephalus platyurus* [153]. All Zn formulations were found to be very toxic with the EC₅₀ values determined for *Vibrio fischeri* being 1,8 mg/L for bulk ZnO, 1,9 mg/L for

nano ZnO, and 1,1 mg/L for ZnSO₄ · 7H₂O, for *Daphnia magna* 8,8 mg/L for bulk ZnO, 3,2 mg/L for nano ZnO and 6,1 mg/L for ZnSO₄ · 7 H₂O, and for *Thamnocephalus platyurus* 0,24 mg/L for bulk ZnO, 0,18 mg/L for nano ZnO, and 0,98 mg/L for ZnSO₄ · 7H₂O, respectively. The toxicity was found to be caused by solubilized Zn ions as proved by tests performed with recombinant Zn-sensor bacteria [153].

Miller et al. showed that ZnO nanoparticles significantly inhibited the growth rate of four different phytoplankton species: *Thalassiosira pseudonana*, *Skeletonema marinoi*, *Dunaliella tertiolecta*, and *Isochrysis galbana*. The ZnO particles aggregated rapidly in seawater forming particles with hydrodynamic diameters exceeding 400 nm [154]. The toxicity of ZnO nanoparticles to phytoplankton was likely due to dissolution, release, and uptake of free zinc ions, with specific nanoparticulate effects being difficult to distinguish from effects due to free zinc ions [154], [155].

The toxicity of Zn and other trace metals to phytoplankton moreover depends on the concentration of other limiting trace metals and nutrients [155]. The mechanism of Zn²⁺-ion toxicity in phytoplankton can be explained by an antagonism between the toxic metal, in this case zinc, and the nutrient metal. Excessive free Zn²⁺-ions competitively inhibit manganese uptake, causing Mn deficiency [156]. Zn²⁺ has also been shown to cause increased ATP production in diatoms, which may be linked to increased thiol and glutathione production [157]. Zn-thiol binding may act as a detoxification mechanism, but the energy required for this reaction may result in a decrease of the cell division rate and therefore of the population growth rate [157].

Mortimer and coworkers investigated the toxic effect of ZnO nanoparticles on the model organism protozoa *Tetrahymena thermophila* and reported the EC₅₀ values after 4 h exposure to be 3,7 mg/L for bulk ZnO, 3,9 mg/L for nano ZnO, and 4,9 mg/L for Zn²⁺, respectively. The toxicity of the zinc compounds was found to be about 1.5 times lower after 24 h of exposure as compared with an exposure time of 4 h, probably due to adaptation of the organisms [158].

The dissolution of ZnO nanoparticles and its contribution to the toxicity on ryegrass was also investigated. Zn²⁺ ions were used to compare and verify the root uptake and phytotoxicity of ZnO nanoparticles in these hydroponic culture systems. The root uptake and the phytotoxicity were visualized by light, scanning electron, and transmission electron microscopy. In the presence of ZnO nanoparticles, ryegrass biomass was significantly reduced, root tips were shrunk, and root epidermal and cortical cells highly vacuolated or collapsed. ZnO nanoparticles were found to be able to concentrate in the rhizosphere, to enter the root cells and to inhibit seedling growth of ryegrass. The phytotoxicity of ZnO nanoparticles could not primarily be explained by their dissolution in the bulk nutrient solution or in the rhizosphere [159].

Because of the entry of nanotechnological products to the air, the water, and the ground nanoparticles can be adsorbed by plants and mammalian organisms. Hence, they enter the food chain and can cause possible hazards and risks to animals and humans. The toxicological aspects to animals and human beings will be discussed in the following section.

5 Human health risks of Nanomaterials

As discussed in Section 4, toxic effects of metal oxide nanomaterials to different bacteria, water organisms and plants have been detected. The potential hazardous effect of semiconductor particles employed in photocatalysis will be summarized in this section. Nanoparticles are able to reach the organism via the lung, via the gastrointestinal tract and partly via the skin. Within the organism they can move freely, and should be able to break through the blood-brain barrier and through cell membranes. Via the blood vessels they can reach and eventually accumulate in different organs such as kidney, heart, and liver. In the body nanoparticles are able to interact with different proteins and cell components as described in Section 3. Therefore, various studies have investigated the toxic effect of TiO₂ and ZnO nanoparticles to different cell lines and animals.

Picatonotto et al. investigated the photocatalytic activity of pigments grade titanium dioxide (TiO₂), used, e.g., in sunscreens and observed a photocatalytic degradation of the organic additives of protective creams and the generation of active species inducing the transformation of biological molecules present on the human skin. Some sunscreens have been photo-degraded after UV-irradiation with the potential risk to act as photosensitizers able to initiate harmful reactions for the skin, especially photo-induced mutagenicity [160]. Hidaka et al. examined the fate of DNA, RNA and their corresponding pyrimidine and purine bases in presence of TiO₂ under UV illumination. Although these in vitro studies may not reflect actual in vivo cases it has been demonstrated that illuminated TiO₂ interacts with DNA or RNA components with serious damage to these substrates being possible [161]. Lu and coworkers have demonstrated that UV-illuminated pure anatase TiO₂ as well as the mixed anatase-rutile Evonik-Degussa Aeroxide P25 can promote the protein tyrosine nitration. The latter is known to occur as a result of oxidative and nitrative stress and is directly involved in the onset or the progression of diseases. On the other hand, the photocatalytic effect of rutile nanoparticles resulting in protein tyrosine nitration was reported to be marginal. Considering the existence of nanostructured TiO₂ in the environment and in cosmetics, e.g., sunscreen products along with the high content of nitrite in sweat, UV-exposed skin may be a significant target for the photosensitized damage [162] and for photo-oxidative injuries

[163], [164], [165]. The potential physiological significance of nano TiO₂ induced photocatalytic protein nitration was also demonstrated in mouse skin homogenate. The relationship between photocatalytic protein tyrosine nitration and chronic cutaneous diseases still needs further investigations [162].

In vitro Studies of TiO₂ and ZnO Nanomaterials

The effect of TiO₂ nanoparticles in different sizes, shapes and surface coatings to various cell lines such as HaCaT cells, an immortalized keratinocyte cell line [166], L929 cells, a mouse fibroblast cell line [167], HeLa cells, an ovarian cell line [168], [169], PC-12 cells, a rat pheochromocytoma cell line [162], [170], [171], A-549, a lung cell line [170], [171], NIH-3T3 cells, a mouse fibroblast cell line [170], [171], and to HEP-G2 cells [170], [171] has been investigated in great detail.

First studies investigating the possible cytotoxic effect of TiO₂ nanoparticles to mammalian cells were carried out by Cai and coworkers in 1992 [169]. They observed that HeLa cells showed no more viability after cultivation with photoactivated (10 min/ 550 W Hg lamp) TiO₂ nanoparticles (P25) at a loading of 50 µg/ml (50 ppm), whereas the treatment with TiO₂ particles in the absence of irradiation showed only a slight decrease of the cell viability [169].

PC-12 cells treated with different concentrations (1, 10, 50, 100 µg/mL) of nanostructured TiO₂ (P25) showed a significant decrease of their viability in periods of 6, 12, 24, and 48 h evincing an explicit dose effect and time dependence. It was suggested that TiO₂ nanoparticles induce intracellular accumulation of reactive oxygen species and the apoptosis of PC-12 cells both of which increase with increasing concentration of TiO₂ [172].

Chen and coworkers tested the cytotoxicity of various anatase nanostructures, possessing three different morphological structures to HeLa cells. 0D anatase nanoparticles at a concentration of 125 µg/mL were found to decrease the cell viability to 80 %, whereas the viability of the cells was close to 100 % when adding the 2D and 3D nanostructures at the same concentration.

By irradiating the treated cells with 8 mW/cm² UV light for 1 min their mortality increased. These differences in toxicity may be related to the different uptake ability of unmodified structures of varying geometry into cells [168]. The studies of Jin et al. showed that weakly aggregated anatase nanoparticles with an average size of less than 100 nm induced significant toxicity in L929 cells at concentrations between 30 µg/mL and 600 µg/mL. The cell shape became spherical accompanied with a cell shrinkage as the concentration of TiO₂ nanoparticles increased. The thus treated cells were found to be necrotic and a significant increase in oxidative stress at higher TiO₂ concentrations (>60 µg/mL) was observed [167].

Oxidative stress and the formation of reactive oxygen species (ROS) were shown to be integral parts of the key mechanisms of cellular defense after particle uptake. TiO₂ nanoparticles evidently induce intracellular oxidative stress by disturbing the balance between oxidant and antioxidant processes [173].

However, other studies have shown that TiO₂ nanomaterials do not exhibit any toxic effect to cells [170], [171]. Nine different powders and three different suspensions, all consisting of TiO₂ nanoparticles have been tested concerning their toxicity to A-549, HEP-G2, PC-12, and NIH-3T3 cells. The particles differed in crystal structure, size and BET-surface area. In one of these studies, none of the tested particles showed any toxic effect on the cells in a concentration range between 100 and 1000 ppm, with the mitochondrial activity of the cells being the only parameter that was determined [170]. Another study investigating the toxic effect of suspensions of different TiO₂ nanoparticles showed that some cells are more sensitive to particle exposure than others. The sensitivity of the employed cell lines was determined to be NIH-3T3 > A-549 > PC-12 > HEP-G2, with the NIH-3T3 being the most sensitive cell line. The highest applied TiO₂ loading of 3125 µg/cm² decreases the viability to 33% (NIH-3T3 cells cultivated with TiO₂ particles with a hydrodynamic diameter of 144.0 ± 23.7 nm) in comparison to untreated cells. However, this study also showed that only very high particle doses exceeding 625 µg/cm² yield a significant decrease in viability. No additional decrease in the viability of cells was observed upon UV(A)-illumination [171].

Several studies have investigated the toxic effect of ZnO nanoparticles to MSTO cells and 3T3-cells [174], to SMMC-7721-cells, a hepatocellular cancer cell line [175], to HeLa and L929 cells [176], to PBMC (peripheral blood mononuclear) cells [177], to human skin fibroblasts [178] to primary mouse embryo fibroblasts, and to A-549 cells. Brunner et al. investigated the toxic effect of ZnO nanoparticles to MSTO- and fibroblasts (3T3-cells) [174] and found that the cells were no longer viable after 3 days exposure to zinc oxide (particle diameter 40 nm) at concentrations exceeding 15 ppm. The authors assumed that the toxic effect of the ZnO nanoparticles is related to the solvated Zn²⁺-ions. Furthermore, Lin et al. investigated the toxic effect of ZnO nanoparticles to A-549 cells at two different particle sizes (70 nm and 420 nm). They conclude that the exposure to both sizes of ZnO particles leads to dose- and time-dependent cytotoxicity reflected in oxidative stress, lipid peroxidation, cell membrane damage, and oxidative DNA damage. Neither free Zn²⁺ ions nor metal impurities appear to be major contributors of ROS induction [179] in contrast to the results of Brunner et al. [174].

Decksakulthorn and coworkers determined the IC₅₀ values of ZnO particles (average particle sizes 70 nm) to be 50 ppm for A-549 cells in contrast to the IC₅₀ values of TiO₂

particles (average particle size 50 nm) being around 2700 ppm [178].

Studies of Hanley and coworkers demonstrate that ZnO nanoparticles induce toxicity in a cell-type specific manner that depends on the degree of particle-cellular membrane association, the phagocytic ability, and the inherent cellular capacities for ROS production. Monocytic cells displayed the greatest susceptibility and intracellular ROS production upon exposure to ZnO nanoparticle, followed by NK cells, and by lymphocytes, which displayed the highest resistance. Hanley et al. assumed that ROS formation is the major mechanism of ZnO nanoparticle-induced toxicity, with the generation of ROS and the cytotoxic effect occurring in a particle size-dependent manner, i.e., smaller particles display the greatest effect [177].

When studying ZnO particle sizes ranging from 20-100 nm Li et al. did not find any particle size dependent toxic effect to SMMC-7721 cells [175].

The proliferation activity of L929 and HeLa cells has been found to be strongly deteriorated upon cultivation in the presence different doses of ZnO nanoparticle (20 nm) suspensions after 24 h and 48 h exposure. Concurrently, an increase in necrotic and apoptotic cells was observed after cultivation with ZnO particles [176].

In vivo Studies of TiO₂ and ZnO Nanoparticles

The effect of various TiO₂ particles in mice, rats, and hamsters after oral exposure and instillation [180], [181], [182], [183], [184], [185], [186], injection [187], [188] and inhalation [189], [181], [190], [191] has been studied in detail.

The inhalation studies with rats showed that a possible hazard and an inflammatory response do exist depending upon the particle characteristics. Lee and coworkers exposed rats to TiO₂ by inhalation exposure with concentrations of 0, 10, 50, and 250 mg/m³ for 6 h/day, 5 days/week for 2 years and found no abnormal clinical signs, no body weight changes, and no excess mortality in any exposed group. However, the exposed groups showed slight increases in the incidence of pneumonia, tracheitis, and rhinitis, and obvious hepatic damage and renal lesion in female mice [188]. TiO₂ particles were found to be mainly retained in liver, kidney, spleen and lung [189]. Based on the excessive dust loading and overwhelmed clearance mechanisms in the lungs of rats exposed chronically at 250 mg/m³, the biological relevance of lung tumors to man appears to be negligible. Furthermore, no evidence exists to suggest that TiO₂ causes lung tumors in humans [189], [192].

Warheit et al. showed in their inhalation studies with rats that the composition and the surface treatment of the nanoparticles can influence the toxicity of TiO₂ particles in the lung. TiO₂ formulations containing 7 % Al₂O₃ and 11 % amorphous silica were found to produce adverse lung effects as compared to pure TiO₂ [181].

Toxicological inhalation studies of Hext et al., with rats, mice, and hamsters being exposed to pigment grade TiO₂ and ultrafine TiO₂ (P25) for 6h/day, 5 days/week for 13 weeks at concentrations of 0, 10, 50, and 250 mg/m³ of TiO₂ were performed to investigate whether rats are oversensitive to TiO₂ compared to mice or hamsters. These epidemiology studies have been carried out to investigate whether a link exists between increased incidence of lung cancer and exposure to TiO₂. The results of these studies do, however, not suggest that TiO₂ nanoparticles exhibit any carcinogenic effect on the human lung [190]. In contrast to the study of Hext et al., Bermudez and coworkers reported that TiO₂ exposure to rats in concentrations of 250 mg/m³ can induce epithelial and fibroproliferative lesions as well as alveolar cell metaplasia using the pure rutile form of TiO₂ for their inhalation studies [191].

Biodistribution experiments performed after oral administration of TiO₂ nanoparticles to rats showed that the particles were retained in liver, spleen, kidney, and lung tissue indicating that TiO₂ nanoparticles can be transported to other tissues and organs after uptake by the gastrointestinal tract [180], [184]. Following inhalation exposure the distribution of the particles was found to be the same [189]. Cui et al. reported that liver damage can be caused by oxidative stress after oral exposure of TiO₂ (100 % anatase) nanoparticles to rats [182].

Hohr and coworkers investigated the acute inflammatory response and the cell damage induced by the intratracheal instillation of surface modified (hydrophilic and hydrophobic) fine (diameter 180 nm) and ultrafine (20 ± 5 nm) TiO₂ particles for 16 h at equivalent mass (1 or 6 mg) and surface doses (100, 500, 600 and 3000 cm²) in rats. The results of Hohr et al. suggest that the surface area rather than the hydrophobicity of the surface determines the acute, pulmonary inflammation induced by both fine and ultrafine TiO₂ [183], whereas Warheit et al. postulated in their studies that nanoscale particles do not exhibit any cytotoxicity or inflammation to the lung as compared to larger sized particles of similar chemical composition [186].

After intraperitoneal injection of high doses of TiO₂ in rats (150 mg/kg) the production of ROS (O^{-•} and H₂O₂) occurred in the liver indicating that this organ underwent oxidative stress which was found to be higher for nano anatase TiO₂ than for bulk TiO₂ [188]. Liu et al. observed that there was a serious damage of liver, kidney, and myocardium, an inflammatory response, and a metabolism imbalance of blood sugar and lipid after abdominal cavity injection of high doses of TiO₂ to mice [187].

Acute toxicity to male mice after intratracheal injection of ZnO particles of different sizes (10, 30, 100 nm) at doses between 0.05 and 0.5 g/kg body weight were also observed. Pathological examination showed the accumulation of ZnO particles after intratracheal injection in lung, spleen, pancreas, bone, and liver. At low doses, 30 and 100 nm sized ZnO particles induced a slight damage, whereas 10 nm

sized particles caused a more serious lung and liver damage. At high doses, the 30 nm sized ZnO particles induced worse lung, liver, and pancreas injury [193].

Wang and coworkers investigated the acute oral toxicity of ZnO particles with different sizes (20 nm and 120 nm) at doses of 1, 2, 3, 4 and 5 g/kg body weight. This study showed that following oral administration ZnO was mainly retained in bone, kidney, and pancreas. The results of blood measurements suggest that the increase in blood viscosity could be induced by high doses of 120 nm sized ZnO nanoparticles. For smaller particles (20 nm) lower concentrations caused the same effect [194].

6 Conclusions

Nanostructured materials are widely used in almost every sector of industry with photocatalysis being one important application field. Suitable semiconductors irradiated with UV- or visible light exhibit photocatalytic activity and can therefore be used for various applications, including the treatment of wastewater and the production of self-cleaning surfaces. Because of the common use of these materials it is important to investigate the possible effects of nanoparticulate photocatalysts to the environment and to human health.

Nanotoxicology is an emerging new subdiscipline of nanotechnology focusing on the toxic potential of nanostructured substances. Particularly difficult tasks within the field of nanotoxicology are the definition of the life cycle of these materials, the handling of the huge amount of natural nanoparticles and the lack of any logical labeling of nanomaterials. Nanoparticles exhibit other physical and chemical properties compared to their bulk counterparts. Therefore, a detailed characterization of these materials is certainly required. Shape, size, size distribution, agglomeration/aggregation, stability, solubility in different media, surface chemistry, composition, charge, crystal structure, zeta potential etc. of the particles have to be determined as a basis for the interpretation of the toxicity studies. To assess their environmental hazards the investigation of the toxic effect of nanomaterials to organisms living in water and to bacteria has to be carried out. For the assessment of the possible hazard to human health, in vitro and in vivo studies need to be performed.

The results of a wide selection of publications investigating the toxic effect of TiO₂ and ZnO, i.e., the two most commonly used photocatalysts, to different bacteria, water organisms, algae and plants as well as in vitro and in vivo studies with different sized and shaped TiO₂ and ZnO particles are presented and discussed. This review shows that considerable differences in the published toxicity results do exist which can mainly be attributed to differences in the types of employed TiO₂ and ZnO nanoparticles (size, shape, etc.) and the treatment methods demonstrating the importance of a suitable characterization of the particles.

In conclusion, it is obvious that the toxic effect of TiO₂ nanoparticles can mainly be attributed to the production of reactive oxygen species, whereas the toxic effect caused by ZnO nanoparticles may be attributed to free Zn²⁺ ions.

References

- [1] Feynman R., There's a plenty of room at the bottom. *Engineering and Science*, 23 (1960): 22–36.
- [2] Schmid G., Nanotechnology: Key technology of the new century – Metal nanoparticles as single-electron switches. *Chemie in Unserer Zeit*, 39(1) (2005): 8–15.
- [3] Murray C. B., Kagan C. R., Bawendi M. G., Synthesis and characterization of monodisperse nanocrystals and close-packed nanocrystal assemblies. *Annual Review of Materials Science*, 30 (2000): 545–610.
- [4] Hireman R., From Moore's Law to Intel innovation-prediction to reality. *Intel Magazine*, (2005): 1–9.
- [5] Leung K. C., Mendes P. M., Magonov S. N., Northrop B. H., Kim S., Patel K., Flood A. H., Tseng H. R., Stoddart J. F., Supramolecular self-assembly of dendronized polymers: reversible control of the polymer architectures through acid-base reactions. *J Am Chem Soc*, 128(33) (2006): 10707–15.
- [6] Grevin B., Rannou P., Electrochemistry: arrays of polymer nanowires. *Nat Mater*, 3(8) (2004): 503–4.
- [7] Salata O., Applications of nanoparticles in biology and medicine. *J Nanobiotechnology*, 2(1) (2004): 3.
- [8] Ma J., Wong H. F., Kong L. B., Peng K. W., Biomimetic processing of nanocrystallite bioactive apatite coating on titanium. *Nanotechnology*, 14(6) (2003): 619–623.
- [9] de la Isla A., Brostow W., Bujard B., Estevez M., Rodriguez J. R., Vargas S., Castano V. M., Nanohybrid scratch resistant coatings for teeth and bone viscoelasticity manifested in tribology. *Materials Research Innovations*, 7(2) (2003): 110–114.
- [10] Sanchez-Martin R. M., Alexander L., Muzerelle M., Cardenas-Maestre J. M., Tsakiridis A., Brickman J. M., Bradley M., Microsphere-mediated protein delivery into cells. *Chembiochem*, 10(9) (2009): 1453–6.
- [11] Pantarotto D., Partidos C. D., Hoebeke J., Brown F., Kramer E., Briand J. P., Muller S., Prato M., Bianco A., Immunization with peptide-functionalized carbon nanotubes enhances virus-specific neutralizing antibody responses. *Chemistry & Biology*, 10(10) (2003): 961–966.
- [12] Bruchez M., Moronne M., Gin P., Weiss S., Alivisatos A. P., Semiconductor nanocrystals as fluorescent biological labels. *Science*, 281(5385) (1998): 2013–2016.
- [13] Chan W. C., Nie S., Quantum dot bioconjugates for ultrasensitive nonisotopic detection. *Science*, 281(5385) (1998): 2016–8.
- [14] Wang S. P., Mamedova N., Kotov N. A., Chen W., Studer J., Antigen/antibody immunocomplex from CdTe nanoparticle bioconjugates. *Nano Letters*, 2(8) (2002): 817–822.
- [15] Edelstein R. L., Tamanaha C. R., Sheehan P. E., Miller M. M., Baselt D. R., Whitman L. J., Colton R. J., The BARC biosensor applied to the detection of biological warfare agents. *Biosensors & Bioelectronics*, 14(10-11) (2000): 805–813.

- [16] Nam J. M., Thaxton C. S., Mirkin C. A., Nanoparticle-based bio-bar codes for the ultrasensitive detection of proteins. *Science*, 301(5641) (2003): 1884–1886.
- [17] Molday R. S., MacKenzie D., Immunospecific ferromagnetic iron-dextran reagents for the labeling and magnetic separation of cells. *J Immunol Methods*, 52(3) (1982): 353–67.
- [18] Shinkai M., Yanase M., Suzuki M., Honda H., Wakabayashi T., Yoshida J., Kobayashi T., Intracellular hyperthermia for cancer using magnetite cationic liposomes. *Journal of Magnetism and Magnetic Materials*, 194(1–3) (1999): 176–184.
- [19] Mazzola L., Commercializing nanotechnology. *Nature Biotechnology*, 21(10) (2003): 1137–1143.
- [20] Paull R., Wolfe J., Hebert P., Sinkula M., Investing in nanotechnology. *Nature Biotechnology*, 21(10) (2003): 1144–1147.
- [21] Vasilev K., Cook J., Griesser H. J. Antibacterial surfaces for biomedical devices. *Expert Review of Medical Devices*, 6(5) (2009): 553–567.
- [22] Fujishima A., Rao T. N., Tryk D. A., Titanium dioxide photocatalysis. *Journal of Photochemistry and Photobiology C Photochemistry Reviews*, 1 (2000): 1–21.
- [23] Fox M. A., Dulay M. T., Heterogeneous Photocatalysis. *Chemical Reviews*, 93(1) (1993): 341–357.
- [24] Patchkovskii S., Tse J. S., Yurchenko S. N., Zhechkov L., Heine T., Seifert G., Graphene nanostructures as tunable storage media for molecular hydrogen. *Proceedings of the National Academy of Sciences of the United States of America*, 102(30) (2005): 10439–10444.
- [25] Zhang X. T., Fujishima A., Jin M., Emeline A. V., Murakami T., Double-layered TiO₂-SiO₂ nanostructured films with self-cleaning and antireflective properties. *Journal of Physical Chemistry B*, 110(50) (2006): 25142–25148.
- [26] Hirai S., Takahashi N., Goto T., Lin S., Uemura T., Yu R., Kawada T. Functional food targeting the regulation of obesity-induced inflammatory responses and pathologies. *Mediators Inflamm*, (2010): 367838.
- [27] Malato S., Fernandez-Ibanez P., Maldonado M. I., Blanco J., Gernjak W., Decontamination and disinfection of water by solar photocatalysis: Recent overview and trends. *Catalysis Today*, 147(1) (2009): 1–59.
- [28] Yu K. H., Chen J. H., Enhancing Solar Cell Efficiencies through 1-D Nanostructures. *Nanoscale Research Letters*, 4(1) (2009): 1–10.
- [29] Fujishima A., Zhang X. T., Tryk D. A., TiO₂ photocatalysis and related surface phenomena. *Surface Science Reports*, 63(12) (2008): 515–582.
- [30] Hoffmann M. R., Martin S. T., Choi W. Y., Bahnemann D. W., Environmental Applications of Semiconductor Photocatalysis. *Chemical Reviews*, 95(1) (1995): 69–96.
- [31] Fujishima A., Rao T. N., Tryk D. A., Titanium dioxide photocatalysis. *Journal of Photochemistry and Photobiology C: Photochemistry Reviews*, 1(1) (2000): 1–21.
- [32] Nosaka Y., Daimon T., Nosaka A. Y., Murakami Y., Singlet oxygen formation in photocatalytic TiO₂ aqueous suspension. *Physical Chemistry Chemical Physics*, 6(11) (2004): 2917–2918.
- [33] Tachikawa T., Fujitsuka M., Majima T., Mechanistic insight into the TiO₂ photocatalytic reactions: Design of new photocatalysts. *Journal of Physical Chemistry C*, 111(14) (2007): 5259–5275.
- [34] Kikuchi Y., Sunada K., Iyoda T., Hashimoto K., Fujishima A., Photocatalytic bactericidal effect of TiO₂ thin films: Dynamic view of the active oxygen species responsible for the effect. *Journal of Photochemistry and Photobiology a-Chemistry*, 106(1–3) (1997): 51–56.
- [35] Naito K., Tachikawa T., Fujitsuka M., Majima T., Single-molecule fluorescence imaging of the remote TiO₂ photocatalytic oxidation. *Journal of Physical Chemistry B*, 109(49) (2005): 23138–23140.
- [36] Park J. S., Choi W., Enhanced remote photocatalytic oxidation on surface-fluorinated TiO₂. *Langmuir*, 20(26) (2004): 11523–11527.
- [37] Tatsuma T., Tachibana S., Fujishima A., Remote oxidation of organic compounds by UV-irradiated TiO₂ via the gas phase. *Journal of Physical Chemistry B*, 105(29) (2001): 6987–6992.
- [38] Naito K., Tachikawa T., Cui S. C., Sugimoto A., Fujitsuka M., Majima T., Single-molecule detection of airborne singlet oxygen. *Journal of the American Chemical Society*, 128(51) (2006): 16430–16431.
- [39] Tatsuma T., Tachibana S., Miwa T., Tryk D. A., Fujishima A., Remote bleaching of methylene blue by UV-irradiated TiO₂ in the gas phase. *Journal of Physical Chemistry B*, 103(38) (1999): 8033–8035.
- [40] Lee N. C., Choi W. Y., Solid phase photocatalytic reaction on the soot/TiO₂ interface: The role of migrating O₂ radicals. *Journal of Physical Chemistry B*, 106(45) (2002): 11818–11822.
- [41] Park J. S., Choi W. Y., Remote photocatalytic oxidation mediated by active oxygen species penetrating and diffusing through polymer membrane over surface fluorinated TiO₂. *Chemistry Letters*, 34(12) (2005): 1630–1631.
- [42] Cho S. M., Choi W. Y., Solid-phase photocatalytic degradation of P. V.C-TiO₂ polymer composites. *Journal of Photochemistry and Photobiology a-Chemistry*, 143(2–3) (2001): 221–228.
- [43] Kawahara K., Ohko Y., Tatsuma T., Fujishima A., Surface diffusion behavior of photo-generated active species or holes on TiO₂ photocatalysts. *Physical Chemistry Chemical Physics*, 5(21) (2003): 4764–4766.
- [44] Haick H., Paz Y., “Dark” photocatalysis: The degradation of organic molecules anchored to dark microdomains of titanium dioxide. *Chemphyschem*, 4(6) (2003): 617–620.
- [45] Ishikawa Y., Matsumoto Y., Nishida Y., Taniguchi S., Watanabe J., Surface treatment of silicon carbide using TiO₂(I. V.) photocatalyst. *Journal of the American Chemical Society*, 125(21) (2003): 6558–6562.
- [46] Tatsuma T., Kubo W., Fujishima A., Patterning of solid surfaces by photocatalytic lithography based on the remote oxidation effect of TiO₂. *Langmuir*, 18(25) (2002): 9632–9634.
- [47] Heller A., Chemistry and applications of photocatalytic oxidation of thin organic films. *Accounts of Chemical Research*, 28(12) (1995): 503–508.

- [48] Maeda K., Domen K., Photocatalytic Water Splitting: Recent Progress and Future Challenges. *Journal of Physical Chemistry Letters*, 1(18) (2010): 2655–2661.
- [49] Kudo A., Miseki Y., Heterogeneous photocatalyst materials for water splitting. *Chemical Society Reviews*, 38(1) (2009): 253–278.
- [50] Woodhouse M., Parkinson B. A., Combinatorial approaches for the identification and optimization of oxide semiconductors for efficient solar photoelectrolysis. *Chemical Society Reviews*, 38(1) (2009): 197–210.
- [51] Garnweitner G., Grote C., In situ investigation of molecular kinetics and particle formation of water-dispersible titania nanocrystals. *Phys Chem Chem Phys*, 11(19) (2009): 3767–74.
- [52] Zhang Z. B., Wang C. C., Zakaria R., Ying J. Y., Role of particle size in nanocrystalline TiO₂-based photocatalysts. *Journal of Physical Chemistry B*, 102(52) (1998): 10871–10878.
- [53] Wang C., Ao Y. H., Wang P. F., Zhang S. H., Qian J., Hou J., A simple method for large-scale preparation of ZnS nanoribbon film and its photocatalytic activity for dye degradation. *Applied Surface Science*, 256(13) (2010): 4125–4128.
- [54] Harris C., Kamat P. V., Photocatalysis with CdSe Nanoparticles in Confined Media: Mapping Charge Transfer Events in the Subpicosecond to Second Timescales. *A. C.S Nano*, 3(3) (2009): 682–690.
- [55] Honda H., Ishizaki A., Soma R., Hashimoto K., Fujishima A., Application of photocatalytic reactions caused by TiO₂ film to improve the maintenance factor of lighting systems. *Journal of the Illuminating Engineering Society*, 27(1) (1998): 42–+.
- [56] Cassar L., Photocatalysis of cementitious materials: Clean buildings and clean air. *Mrs Bulletin*, 29(5) (2004): 328–331.
- [57] Zhang X. T., Sato O., Taguchi M., Einaga Y., Murakami T., Fujishima A., Self-cleaning particle coating with antireflection properties. *Chemistry of Materials*, 17(3) (2005): 696–700.
- [58] Tung W. S., Daoud W. A., Photocatalytic self-cleaning keratins: A feasibility study. *Acta Biomaterialia*, 5(1) (2009): 50–56.
- [59] Rupp F., Haupt M., Klostermann H., Kim H. S., Eichler M., Peetsch A., Scheideler L., Doering C., Oehr C., Wendel H. P., Sinn S., Decker E., von Ohle C., Geis-Gerstorfer J., Multifunctional nature of UV-irradiated nanocrystalline anatase thin films for biomedical applications. *Acta Biomater*, 6(12): 4566–77.
- [60] Yao Y., Ohko Y., Sekiguchi Y., Fujishima A., Kubota Y., Self-sterilization using silicone catheters coated with Ag and TiO₂ nanocomposite thin film. *Journal of Biomedical Materials Research Part B-Applied Biomaterials*, 85B(2) (2008): 453–460.
- [61] Nakamura H., Tanaka M., Shinohara S., Gotoh M., Karube I., Development of a self-sterilizing lancet coated with a titanium dioxide photocatalytic nano-layer for self-monitoring of blood glucose. *Biosens Bioelectron*, 22(9–10) (2007): 1920–5.
- [62] Pichat P., Disdier J., Hoang-Van C., Mas D., Goutailler G., Gaysse C., Purification/deodorization of indoor air and gaseous effluents by TiO₂ photocatalysis. *Catalysis Today*, 63(2–4) (2000): 363–369.
- [63] Ao C. H., Lee S. C., Enhancement effect of TiO₂ immobilized on activated carbon filter for the photodegradation of pollutants at typical indoor air level. *Applied Catalysis B-Environmental*, 44(3) (2003): 191–205.
- [64] Ao C. H., Lee S. C., Combination effect of activated carbon with TiO₂ for the photodegradation of binary pollutants at typical indoor air level. *Journal of Photochemistry and Photobiology A – Chemistry*, 161(2–3) (2004): 131–140.
- [65] Kim J. H., Seo G., Cho D. L., Choi B. C., Kim J. B., Park H. J., Kim M. W., Song S. J., Kim G. J., Kato S., Development of air purification device through application of thin-film photocatalyst. *Catalysis Today*, 111(3–4) (2006): 271–274.
- [66] Grinshpun S. A., Adhikari A., Honda T., Kim K. Y., Toivola M., Rao K. S.R., Reponen T., Control of aerosol contaminants in indoor air: Combining the particle concentration reduction with microbial inactivation. *Environmental Science & Technology*, 41(2) (2007): 606–612.
- [67] Herrmann J. M., Heterogeneous photocatalysis: State of the art and present applications. *Topics in Catalysis*, 34(1–4) (2005): 49–65.
- [68] Oller I., Gernjak W., Maldonado M. I., Perez-Estrada L. A., Sanchez-Perez J. A., Malato S., Solar photocatalytic degradation of some hazardous water-soluble pesticides at pilot-plant scale. *Journal of Hazardous Materials*, 138(3) (2006): 507–517.
- [69] Perez M. H., Penuela G., Maldonado M. I., Malato O., Fernandez-Ibanez P., Oller I., Gernjak W., Malato S., Degradation of pesticides in water using solar advanced oxidation processes. *Applied Catalysis B-Environmental*, 64(3–4) (2006): 272–281.
- [70] Kositzki M., Poullos I., Malato S., Caceres J., Campos A., Solar photocatalytic treatment of synthetic municipal wastewater. *Water Research*, 38(5) (2004): 1147–1154.
- [71] Maldonado M. I., Passarinho P. C., Oller I., Gernjak W., Fernandez P., Blanco J., Malato S., Photocatalytic degradation of E. U. priority substances: A comparison between TiO₂ and Fenton plus photo-Fenton in a solar pilot plant. *Journal of Photochemistry and Photobiology A – Chemistry*, 185(2–3) (2007): 354–363.
- [72] Maynard A. D., Aitken R. J., Butz T., Colvin V., Donaldson K., Oberdorster G., Philbert M. A., Ryan J., Seaton A., Stone V., Tinkle S. S., Tran L., Walker N. J., Warheit D. B., Safe handling of nanotechnology. *Nature*, 444(7117) (2006): 267–269.
- [73] Tsuji J. S., Maynard A. D., Howard P. C., James J. T., Lam C. W., Warheit D. B., Santamaria A. B., Research strategies for safety evaluation of nanomaterials, part IV: Risk assessment of nanoparticles. *Toxicological Sciences*, 89(1) (2006): 42–50.
- [74] Oberdorster G. Safety assessment for nanotechnology and nanomedicine: concepts of nanotoxicology. *J Intern Med*, 267(1): 89–105.
- [75] Oberdorster G., Oberdorster E., Oberdorster J., Nanotoxicology: an emerging discipline evolving from studies of ultrafine particles. *Environ Health Perspect*, 113(7) (2005): 823–39.

- [76] Oberdorster G., Stone V., Donaldson K., Toxicology of nanoparticles: A historical perspective. *Nanotoxicology*, 1(1) (2007): 2–25.
- [77] Maynard A. D., Aitken R. J., Assessing exposure to airborne nanomaterials: Current abilities and future requirements. *Nanotoxicology*, 1(1) (2007): 26–41.
- [78] Seipenbusch M., Binder A., Kasper G., Temporal evolution of nanoparticle aerosols in workplace exposure. *Ann Occup Hyg*, 52(8) (2008): 707–16.
- [79] Fujitani Y., Kobayashi T., Arashidani K., Kunugita N., Suemura K., Measurement of the physical properties of aerosols in a fullerene factory for inhalation exposure assessment. *J Occup Environ Hyg*, 5(6) (2008): 380–9.
- [80] Han J. H., Lee E. J., Lee J. H., So K. P., Lee Y. H., Bae G. N., Lee S. B., Ji J. H., Cho M. H., Yu I. J., Monitoring multiwalled carbon nanotube exposure in carbon nanotube research facility. *Inhalation Toxicology*, 20(8) (2008): 741–749.
- [81] Mitsakou C., Helmis C., Housiadas C., Eulerian modelling of lung deposition with sectional representation of aerosol dynamics. *Journal of Aerosol Science*, 36(1) (2005): 75–94.
- [82] Aitken R., RS Report Nanoscience and nanotechnology, (2004).
- [83] Gwinn M. R., Tran L., Risk management of nanomaterials. *Wiley Interdisciplinary Reviews-Nanomedicine and Nanobiotechnology*, 2(2) (2010): 130–137.
- [84] Teeguarden J. G., Hinderliter P. M., Orr G., Thrall B. D., Pounds J. G., Particokinetics in vitro: Dosimetry considerations for in vitro nanoparticle toxicology assessments (vol 95, pg 300, 2007). *Toxicological Sciences*, 97(2) (2007): 614–614.
- [85] Lam C. W., James J. T., McCluskey R., Hunter R. L., Pulmonary toxicity of single-wall carbon nanotubes in mice 7 and 90 days after intratracheal instillation. *Toxicological Sciences*, 77(1) (2004): 126–134.
- [86] Warheit D. B., Laurence B. R., Reed K. L., Roach D. H., Reynolds G. A. M., Webb T. R., Comparative pulmonary toxicity assessment of single-wall carbon nanotubes in rats. *Toxicological Sciences*, 77(1) (2004): 117–125.
- [87] Helland A., Wick P., Koehler A., Schmid K., Som C., Reviewing the environmental and human health knowledge base of carbon nanotubes. *Environmental Health Perspectives*, 115(8) (2007): 1125–1131.
- [88] Oberdorster G., Ferin J., Gelein R., Soderholm S. C., Finkelstein J., Role of the alveolar macrophage in lung injury: studies with ultrafine particles. *Environ Health Perspect*, 97 (1992):193–9.
- [89] Donaldson K., Tran L., Jimenez L. A., Duffin R., Newby D. E., Mills N., MacNee W., Stone V., Combustion-derived nanoparticles (2005): a review of their toxicology following inhalation exposure. *Part Fibre Toxicol*, 2(10).
- [90] Dybdahl M., Risom L., Bornholdt J., Autrup H., Loft S., Wallin H., Inflammatory and genotoxic effects of diesel particles in vitro and in vivo. *Mutation Research-Genetic Toxicology and Environmental Mutagenesis*, 562(1–2) (2004): 119–131.
- [91] Hirano S., Furuyama A., Koike E., Kobayashi T., Oxidative-stress potency of organic extracts of diesel exhaust and urban fine particles in rat heart microvessel endothelial cells. *Toxicology*, 187(2–3) (2003): 161–170.
- [92] McNeilly J. D., Heal M. R., Beverland I. J., Howe A., Gibson M. D., Hibbs L. R., MacNee W., Donaldson K., Soluble transition metals cause the pro-inflammatory effects of welding fumes in vitro. *Toxicology and Applied Pharmacology*, 196(1) (2004): 95–107.
- [93] Renwick L. C., Brown D., Clouter A., Donaldson K., Increased inflammation and altered macrophage chemotactic responses caused by two ultrafine particle types. *Occupational and Environmental Medicine*, 61(5) (2004): 442–447.
- [94] Gilmour M. I., O'Connor S., Dick C. A.J., Miller C. A., Linak W. P., Differential pulmonary inflammation and in vitro cytotoxicity of size-fractionated fly ash particles from pulverized coal combustion. *Journal of the Air & Waste Management Association*, 54(3) (2004): 286–295.
- [95] Lockman P. R., Koziara J. M., Mumper R. J., Allen D. D., Nanoparticle surface charges alter blood-brain barrier integrity and permeability. *J Drug Target*, 12(9–10) (2004): 635–41.
- [96] Lademann J., Weigmann H., Rickmeyer C., Barthelmes H., Schaefer H., Mueller G., Sterry W., Penetration of titanium dioxide microparticles in a sunscreen formulation into the horny layer and the follicular orifice. *Skin Pharmacol Appl Skin Physiol*, 12(5) (1999): 247–56.
- [97] Alvarez-Roman R., Naik A., Kalia Y., Guy R. H., Fessi H., Skin penetration and distribution of polymeric nanoparticles. *Journal of Controlled Release*, 99(1) (2004): 53–62.
- [98] Zvyagin A. V., Zhao X., Gierden A., Sanchez W., Ross J. A., Roberts M. S., Imaging of zinc oxide nanoparticle penetration in human skin in vitro and in vivo. *J Biomed Opt*, 13(6) (2008): 064031.
- [99] Cross S. E., Innes B., Roberts M. S., Tsuzuki T., Robertson T. A., McCormick P., Human skin penetration of sunscreen nanoparticles: In-vitro assessment of a novel micronized zinc oxide formulation. *Skin Pharmacology and Physiology*, 20(3) (2007): 148–154.
- [100] Mortensen L. J., Oberdorster G., Pentland A. P., Delouise L. A., In vivo skin penetration of quantum dot nanoparticles in the murine model: the effect of UVR. *Nano Lett*, 8(9) (2008): 2779–87.
- [101] Ito A., Shinkai M., Honda H., Kobayashi T., Medical application of functionalized magnetic nanoparticles. *Journal of Bioscience and Bioengineering*, 100(1) (2005): 1–11.
- [102] Gopee N. V., Roberts D. W., Webb P., Cozart C. R., Sintonen P. H., Warbritton A. R., Yu W. W., Colvin V. L., Walker N. J., Howard P. C., Migration of intradermally injected quantum dots to sentinel organs in mice. *Toxicol Sci*, 98(1) (2007): 249–57.
- [103] Chithrani B. D., Ghazani A. A., Chan W. C., Determining the size and shape dependence of gold nanoparticle uptake into mammalian cells. *Nano Lett*, 6(4) (2006): 662–8.
- [104] Choi H. S., Liu W., Misra P., Tanaka E., Zimmer J. P., Ipe B. I., Bawendi M. G., Frangioni J. V., Renal clearance of quantum dots. *Nature Biotechnology*, 25(10) (2007): 1165–1170.
- [105] Fischer H. C., Liu L. C., Pang K. S., Chan W. C.W., Pharmacokinetics of nanoscale quantum dots: In vivo distribution, sequestration, and clearance in the rat. *Advanced Functional Materials*, 16(10) (2006): 1299–1305.

- [106] Nel A., Xia T., Madler L., Li N., Toxic potential of materials at the nanolevel. *Science*, 311(5761) (2006): 622–627.
- [107] Sayes C. M., Gobin A. M., Ausman K. D., Mendez J., West J. L., Colvin V. L., Nano-C-60 cytotoxicity is due to lipid peroxidation. *Biomaterials*, 26(36) (2005): 7587–7595.
- [108] Ipe B. I., Lehnig M., Niemeyer C. M., On the generation of free radical species from quantum dots. *Small*, 1(7) (2005): 706–709.
- [109] Lovric J., Cho S. J., Winnik F. M., Maysinger D., Unmodified cadmium telluride quantum dots induce reactive oxygen species formation leading to multiple organelle damage and cell death. *Chemistry & Biology*, 12(11) (2005): 1227–1234.
- [110] Tsay J. M., Michalet X., New light on quantum dot cytotoxicity. *Chemistry & Biology*, 12(11) (2005): 1159–1161.
- [111] Klein J., Probing the interactions of proteins and nanoparticles. *Proceedings of the National Academy of Sciences of the United States of America*, 104(7) (2007): 2029–2030.
- [112] Cedervall T., Lynch I., Lindman S., Berggard T., Thulin E., Nilsson H., Dawson K. A., Linse S., Understanding the nanoparticle-protein corona using methods to quantify exchange rates and affinities of proteins for nanoparticles. *Proceedings of the National Academy of Sciences of the United States of America*, 104(7) (2007): 2050–2055.
- [113] Petkovic J., Zegura B., Stevanovic M., Drnovsek N., Uskokovic D., Novak S., Filipic M., DNA damage and alterations in expression of DNA damage responsive genes induced by TiO₂ nanoparticles in human hepatoma HepG2 cells. *Nanotoxicology*,
- [114] Ku S., Yan F., Wang Y., Sun Y., Yang N., Ye L. The blood-brain barrier penetration and distribution of P. E. Gylated fluorescein-doped magnetic silica nanoparticles in rat brain. *Biochem Biophys Res Commun*, 394(4): 871–6.
- [115] Meißner T., Potthoff A., Richter V., Suspension characterization as important key for toxicological investigations. *Journal of Physics*, Conference Series 170 (Nanosafe 2008).
- [116] Allouni Z. E., Cimpan M. R., Hol P. J., Skodvin T., Gjerdet N. R. **2009**, Agglomeration and sedimentation of TiO₂ nanoparticles in cell culture medium. *Colloids and Surfaces B-Biointerfaces*, 68(1) (2009): 83–87.
- [117] Buford M. C., Hamilton R. F., Jr., Holian A., A comparison of dispersing media for various engineered carbon nanoparticles. *Part Fibre Toxicol*, 4: (2007)6.
- [118] Kuhnel D., Busch W., Meissner T., Springer A., Potthoff A., Richter V., Gelinsky M., Scholz S., Schirmer K., Agglomeration of tungsten carbide nanoparticles in exposure medium does not prevent uptake and toxicity toward a rainbow trout gill cell line. *Aquat Toxicol*, 93(2–3) (2009): 91–9.
- [119] Chen Z. P., Xu R. Z., Zhang Y., Gu N., Effects of Proteins from Culture Medium on Surface Property of Silanes-Functionalized Magnetic Nanoparticles. *Nanoscale Res Lett*, 4(3) (2008): 204–209.
- [120] Ji Z., Jin X., George S., Xia T., Meng H., Wang X., Suarez E., Zhang H., Hoek E. M., Godwin H., Nel A. E., Zink J. I., Dispersion and stability optimization of TiO₂ nanoparticles in cell culture media. *Environ Sci Technol*, 44(19): 7309–14.
- [121] Kan A. T., Tomson M. B., Ground-Water Transport of Hydrophobic Organic-Compounds in the Presence of Dissolved Organic-Matter. *Environmental Toxicology and Chemistry*, 9(3) (1990): 253–263.
- [122] Jacoby W. A., Maness P. C., Wolfrum E. J., Blake D. M., Fennell J. A., Mineralization of bacterial cell mass on a photocatalytic surface in air. *Environmental Science & Technology*, 32(17) (1998): 2650–2653.
- [123] Matsunaga T., Tomoda R., Nakajima T., Wake H., Photoelectrochemical Sterilization of Microbial-Cells by Semiconductor Powders. *Fems Microbiology Letters*, 29(1–2) (1985): 211–214.
- [124] Coleman H. M., Marquis C. P., Scott J. A., Chin S. S., Amal R., Bactericidal effects of titanium dioxide-based photocatalysts. *Chemical Engineering Journal*, 113(1) (2005): 55–63.
- [125] Srinivasan C., Somasundaram N., Bactericidal and detoxification effects of irradiated semiconductor catalyst, TiO₂. *Current Science*, 85(10) (2003): 1431–1438.
- [126] Chen F. N., Yang X. D., Xu F. F., Wu Q., Zhang Y. P., Correlation of Photocatalytic Bactericidal Effect and Organic Matter Degradation of TiO₂ Part I: Observation of Phenomena. *Environmental Science & Technology*, 43(4) (2009): 1180–1184.
- [127] Liu H. L., Yang T. C.K., Photocatalytic inactivation of *Escherichia coli* and *Lactobacillus helveticus* by ZnO and TiO₂ activated with ultraviolet light. *Process Biochemistry*, 39(4) (2003): 475–481.
- [128] Armon R., Laot N., Neeman I., Photocatalytic inactivation of different bacteria and bacteriophages in drinking water at different TiO₂ concentration with or without exposure to O₂. *Journal of Advanced Oxidation*, 3 (1998): 145–150.
- [129] Biguzzi M., Shama G., Effect of Titanium-Dioxide Concentration on the Survival of *Pseudomonas-Stutzeri* during Irradiation with near-Ultraviolet Light. *Letters in Applied Microbiology*, 19(6) (1994): 458–460.
- [130] Pham H. N., McDowell T., Wilkins E., Photocatalytically-Mediated Disinfection of Water Using TiO₂ as a Catalyst and Spore-Forming *Bacillus-Pumilus* as a Model. *Journal of Environmental Science and Health Part a-Environmental Science and Engineering & Toxic and Hazardous Substance Control*, 30(3) (1995): 627–636.
- [131] Kuhn K. P., Chaberny I. F., Massholder K., Stickler M., Benz V. W., Sonntag H. G., Erdinger L., Disinfection of surfaces by photocatalytic oxidation with titanium dioxide and UVA light. *Chemosphere*, 53(1) (2003): 71–7.
- [132] Gogniat G., Thyssen M., Denis M., Pulgarin C., Dukan S., The bactericidal effect of TiO₂ photocatalysis involves adsorption onto catalyst and the loss of membrane integrity. *Fems Microbiology Letters*, 258(1) (2006): 18–24.
- [133] Yao K. S., Wang D. Y., Chang C. Y., Weng K. W., Yang L. Y., Lee S. J., Cheng T. C., Hwang C. C., Photocatalytic disinfection of phytopathogenic bacteria by dye-sensitized TiO₂ thin film activated by visible light. *Surface & Coatings Technology*, 202(4–7) (2007): 1329–1332.
- [134] Yao K. S., Wang D. Y., Ho W. Y., Yan J. J., Tzeng K. C., Photocatalytic bactericidal effect of TiO₂ thin film on plant pathogens. *Surface & Coatings Technology*, 201(15) (2007): 6886–6888.
- [135] Huang Z. B., Zheng X., Yan D. H., Yin G. F., Liao X. M., Kang Y. Q., Yao Y. D., Huang D., Hao B. Q., Toxicological

- effect of ZnO nanoparticles based on bacteria. *Langmuir*, 24(8) (2008): 4140–4144.
- [136] Adams L. K., Lyon D. Y., Alvarez P. J.J., Comparative ecotoxicity of nanoscale TiO₂, SiO₂, and ZnO water suspensions. *Water Research*, 40(19) (2006): 3527–3532.
- [137] Zhang L. L., Jiang Y. H., Ding Y. L., Povey M., York D., Investigation into the antibacterial behaviour of suspensions of ZnO nanoparticles (ZnO nanofluids). *Journal of Nanoparticle Research*, 9(3) (2007): 479–489.
- [138] Roselli M., Finamore A., Garaguso I., Britti M. S., Mengheri E., Zinc oxide protects cultured enterocytes from the damage induced by *Escherichia coli*. *Journal of Nutrition*, 133(12) (2003): 4077–4082.
- [139] Jones N., Ray B., Ranjit K. T., Manna A. C., Antibacterial activity of ZnO nanoparticle suspensions on a broad spectrum of microorganisms. *Fems Microbiology Letters*, 279(1) (2008): 71–76.
- [140] Sunada K., Kikuchi Y., Hashimoto K., Fujishima A., Bactericidal and detoxification effects of TiO₂ thin film photocatalysts. *Environmental Science & Technology*, 32(5) (1998): 726–728.
- [141] Sunada K., Watanabe T., Hashimoto K., Studies on photokilling of bacteria on TiO₂ thin film. *Journal of Photochemistry and Photobiology a-Chemistry*, 156(1–3) (2003): 227–233.
- [142] Brayner R., Ferrari-Iliou R., Brivois N., Djediat S., Benedetti M. F., Fievet F., Toxicological impact studies based on *Escherichia coli* bacteria in ultrafine ZnO nanoparticles colloidal medium. *Nano Letters*, 6(4) (2006): 866–870.
- [143] Gaballa A., Helmann J. D., Identification of a zinc-specific metalloregulatory protein, Zur, controlling zinc transport operons in *Bacillus subtilis*. *J Bacteriol*, 180(22) (1998): 5815–21.
- [144] Lindsay J. A., Foster S. J., zur: a Zn(2+)-responsive regulatory element of *Staphylococcus aureus*. *Microbiology*, 147(Pt 5) (2001): 1259–66.
- [145] Hund-Rinke K., Simon M., Ecotoxic effect of photocatalytic active nanoparticles (TiO₂) on algae and daphnids. *Environ Sci Pollut Res Int*, 13(4) (2006): 225–32.
- [146] Lovern S. B., Klaper R., *Daphnia magna* mortality when exposed to titanium dioxide and fullerene (C60) nanoparticles. *Environ Toxicol Chem*, 25(4) (2006): 1132–7.
- [147] Velzeboer I., Hendriks A. J., Ragas A. M., Van de Meent D., Aquatic ecotoxicity tests of some nanomaterials. *Environ Toxicol Chem*, 27(9) (2008): 1942–7.
- [148] Ruiz M. J., Lopez-Jaramillo L., Redondo M. J., Font G., Toxicity assessment of pesticides using the microtox test: application to environmental samples. *Bull Environ Contam Toxicol*, 59(4) (1997): 619–25.
- [149] Pena-Vazquez E., Perez-Conde C., Costas E., Moreno-Bondi M. C. Development of a microalgal P. A.M test method for Cu(II) in waters: comparison of using spectrofluorometry. *Ecotoxicology*, 19(6): 1059–65.
- [150] Pieters B., Bosman-Meijerman D., Steenbergen E., van de Brandhof E.-J., van Beelen P., van de Grinten E., Verweij W., Kraak M., Ecological quality assessment of Dutch surface waters using a new bioassays with the cladoceran *Chydorus sphaericus*. *Proc Neth Entomol Soc Meet*, 19 (2008): 157–164.
- [151] Schmitt H., Martinali B., Van Beelen P., Seinen W., On the limits of toxicant-induced tolerance testing: cotolerance and response variation of antibiotic effects. *Environ Toxicol Chem*, 25(7) (2006): 1961–8.
- [152] Aruoja V., Dubourguier H. C., Kasemets K., Kahru A., Toxicity of nanoparticles of CuO, ZnO and TiO₂ to microalgae *Pseudokirchneriella subcapitata*. *Sci Total Environ*, 407(4) (2009): 1461–8.
- [153] Heinlaan M., Ivask A., Blinova I., Dubourguier H. C., Kahru A., Toxicity of nanosized and bulk ZnO, CuO and TiO₂ to bacteria *Vibrio fischeri* and crustaceans *Daphnia magna* and *Thamnocephalus platyurus*. *Chemosphere*, 71(7) (2008): 1308–16.
- [154] Miller R. J., Lenihan H. S., Muller E. B., Tseng N., Hanna S. K., Keller A. A., Impacts of metal oxide nanoparticles on marine phytoplankton. *Environ Sci Technol*, 44(19): 7329–34.
- [155] Sunda W. G., Price N. M., Morel F. M. M., Trace metal ion buffers and their use in culture studies. *Algal Culturing Techniques* (2005), 35–64.
- [156] Sunda W. G., Huntsman S. A., Antagonisms between cadmium and zinc toxicity and manganese limitation in a coastal diatom. *Limnology and Oceanography*, 41(3) (1996): 373–387.
- [157] Stauber J. L., Florence T. M., Mechanism of Toxicity of Zinc to the Marine Diatom *Nitzschia-Closterium*. *Marine Biology*, 105(3) (1990): 519–524.
- [158] Mortimer M., Kasemets K., Kahru A., Toxicity of ZnO and CuO nanoparticles to ciliated protozoa *Tetrahymena thermophila*. *Toxicology*, 269(2–3) (2010): 182–189.
- [159] Lin D. H., Xing B. S., Root uptake and phytotoxicity of ZnO nanoparticles. *Environmental Science & Technology*, 42(15) (2008): 5580–5585.
- [160] Picatonotto T., Vione D., Carlotti M. E., Gallarate M., Photocatalytic activity of inorganic sunscreens. *Journal of Dispersion Science and Technology*, 22(4) (2001): 381–386.
- [161] Hidaka H., Horikoshi S., Serpone N., Knowland J., In vitro photochemical damage to DNA, RNA and their bases by an inorganic sunscreen agent on exposure to UVA and UVB radiation. *Journal of Photochemistry and Photobiology a-Chemistry*, 111(1–3) (1997): 205–213.
- [162] Lu N. H., Zhu Z. N., Zhao X. Q., Tao R., Yang X. L., Gao Z. H., Nano titanium dioxide photocatalytic protein tyrosine nitration: A potential hazard of TiO₂ on skin. *Biochemical and Biophysical Research Communications*, 370(4) (2008): 675–680.
- [163] Herrling T., Jung K., Fuchs J., Measurements of UV-generated free radicals/reactive oxygen species (ROS) in skin. *Spectrochimica Acta Part A – Molecular and Biomolecular Spectroscopy*, 63(4) (2006): 840–845.
- [164] Brezova V., Gabcova S., Dvoranova D., Stasko A., Reactive oxygen species produced upon photoexcitation of sunscreens containing titanium dioxide (an EPR study). *Journal of Photochemistry and Photobiology B – Biology*, 79(2) (2005): 121–134.
- [165] Dunford R., Salinaro A., Cai L. Z., Serpone N., Horikoshi S., Hidaka H., Knowland J., Chemical oxidation and DNA damage catalysed by inorganic sunscreen ingredients. *Febs Letters*, 418(1–2) (1997): 87–90.

- [166] Jin C., Tang Y., Yang F. G., Li X. L., Xu S., Fan X. Y., Huang Y. Y., Yang Y. J. Cellular Toxicity of TiO₂ Nanoparticles in Anatase and Rutile Crystal Phase. *Biol Trace Elem Res*,
- [167] Jin C. Y., Zhu B. S., Wang X. F., Lu Q. H., Cytotoxicity of titanium dioxide nanoparticles in mouse fibroblast cells. *Chemical Research in Toxicology*, 21(9) (2008): 1871–1877.
- [168] Chen J. Y., Zhou H. J., Santulli A. C., Wong S. S., Evaluating Cytotoxicity and Cellular Uptake from the Presence of Various Processed TiO₂ Nanostructured Morphologies. *Chemical Research in Toxicology*, 23(5) (2010): 871–879.
- [169] Cai R. X., Kubota Y., Shuin T., Sakai H., Hashimoto K., Fujishima A., Induction of Cytotoxicity by Photoexcited TiO₂ Particles. *Cancer Research*, 52(8) (1992): 2346–2348.
- [170] Wagner S., Münzer S., Behrens P., Scheper T., Bahnemann D. W., Kasper C., Cytotoxicity of Titanium and Silicon Dioxide Nanoparticles. *Journal of Physics, Conference Series* 170 (Nanosafe 2008).
- [171] Bloh J. Z., Wagner S., Bahnemann D. W., Scheper T., Kasper C. **2010**, Studies on cytotoxicity of photocatalytic active titanium dioxide nanoparticles. *Chemie Ingenieur Technik*, 82(3) (2009): 335–341.
- [172] Liu S. C., Xu L. J., Zhang T., Ren G. G., Yang Z., Oxidative stress and apoptosis induced by nanosized titanium dioxide in PC12 cells. *Toxicology*, 267(1–3) (2010): 172–177.
- [173] Gurr J. R., Wang A. S. S., Chen C. H., Jan K. Y., Ultrafine titanium dioxide particles in the absence of photoactivation can induce oxidative damage to human bronchial epithelial cells. *Toxicology*, 213(1–2) (2005): 66–73.
- [174] Brunner T. J., Wick P., Manser P., Spohn P., Grass R. N., Limbach L. K., Bruinink A., Stark W. J., In vitro cytotoxicity of oxide nanoparticles: Comparison to asbestos, silica, and the effect of particle solubility. *Environmental Science & Technology*, 40(14) (2006): 4374–4381.
- [175] Li J. Y., Guo D. D., Wang X. M., Wang H. P., Jiang H., Chen B. A., The Photodynamic Effect of Different Size ZnO Nanoparticles on Cancer Cell Proliferation In Vitro. *Nanoscale Research Letters*, 5(6) (2010): 1063–1071.
- [176] Zheng Y. F., Li R. Z., Wang Y. D., In Vitro and in Vivo Biocompatibility Studies of ZnO Nanoparticles. *International Journal of Modern Physics B*, 23(6–7) (2009): 1566–1571.
- [177] Hanley C., Thurber A., Hanna C., Punnoose A., Zhang J. H., Wingett D. G., The Influences of Cell Type and ZnO Nanoparticle Size on Immune Cell Cytotoxicity and Cytokine Induction. *Nanoscale Research Letters*, 4(12) (2009): 1409–1420.
- [178] Decksakulthorn F., Hayes A., Bakand S., Joeng L., Winder C., In vitro cytotoxicity assessment of selected nanoparticles using human skin fibroblasts. *AATEX, Special Issue* (2007) (2007): 397–400.
- [179] Lin W. S., Xu Y., Huang C. C., Ma Y. F., Shannon K. B., Chen D. R., Huang Y. W., Toxicity of nano- and micro-sized ZnO particles in human lung epithelial cells. *Journal of Nanoparticle Research*, 11(1) (2009): 25–39.
- [180] Wang J., Zhou G., Chen C., Yu H., Wang T., Ma Y., Jia G., Gao Y., Li B., Sun J., Li Y., Jiao F., Zhao Y., Chai Z., Acute toxicity and biodistribution of different sized titanium dioxide particles in mice after oral administration. *Toxicol Lett*, 168(2) (2007): 176–85.
- [181] Warheit D. B., Brock W. J., Lee K. P., Webb T. R., Reed K. L., Comparative pulmonary toxicity inhalation and instillation studies with different TiO₂ particle formulations: impact of surface treatments on particle toxicity. *Toxicol Sci*, 88(2) (2005): 514–24.
- [182] Cui Y., Gong X., Duan Y., Li N., Hu R., Liu H., Hong M., Zhou M., Wang L., Wang H., Hong F. Hepatocyte apoptosis and its molecular mechanisms in mice caused by titanium dioxide nanoparticles. *J Hazard Mater*, 183(1–3): 874–80.
- [183] Hohr D., Steinfartz Y., Schins R. P., Knaapen A. M., Martra G., Fubini B., Borm P. J., The surface area rather than the surface coating determines the acute inflammatory response after instillation of fine and ultrafine TiO₂ in the rat. *Int J Hyg Environ Health*, 205(3) (2002): 239–44.
- [184] Jani P. U., Mccarthy D. E., Florence A. T., Titanium-Dioxide (Rutile) Particle Uptake from the Rat GI Tract and Translocation to Systemic Organs after Oral-Administration. *International Journal of Pharmaceutics*, 105(2) (1994): 157–168.
- [185] Duan Y., Liu J., Ma L., Li N., Liu H., Wang J., Zheng L., Liu C., Wang X., Zhao X., Yan J., Wang S., Wang H., Zhang X., Hong F. Toxicological characteristics of nanoparticulate anatase titanium dioxide in mice. *Biomaterials*, 31(5): 894–9.
- [186] Warheit D. B., Webb T. R., Sayes C. M., Colvin V. L., Reed K. L., Pulmonary instillation studies with nanoscale TiO₂ rods and dots in rats: toxicity is not dependent upon particle size and surface area. *Toxicol Sci*, 91(1) (2006): 227–36.
- [187] Liu H., Ma L., Zhao J., Liu J., Yan J., Ruan J., Hong F., Biochemical toxicity of nano-anatase TiO₂ particles in mice. *Biol Trace Elem Res*, 129(1–3) (2009): 170–80.
- [188] Liu H., Ma L., Liu J., Zhao J., Yan J., Hong F., Toxicity of nano-anatase TiO₂ to mice: Liver injury oxidative stress. *Toxicological and Environmental Chemistry*, 92 (2010): 175–186.
- [189] Lee K. P., Trochimowicz H. J., Reinhardt C. F., Pulmonary response of rats exposed to titanium dioxide (TiO₂) by inhalation for two years. *Toxicol Appl Pharmacol*, 79(2) (1985): 179–92.
- [190] Hext P. M., Tomenson J. A., Thompson P., Titanium dioxide: inhalation toxicology and epidemiology. *Ann Occup Hyg*, 49(6) (2005): 461–72.
- [191] Bermudez E., Mangum J. B., Asgharian B., Wong B. A., Reverdy E. E., Janszen D. B., Hext P. M., Warheit D. B., Everitt J. I., Long-term pulmonary responses of three laboratory rodent species to subchronic inhalation of pigmented titanium dioxide particles. *Toxicol Sci*, 70(1) (2002): 86–97.
- [192] Uragoda C. G., Pinto M. R., An investigation into the health of workers in an ilmenite extracting plant. *Med J Aust*, 1(4) (1972): 167–9.
- [193] Song W., Zhang J., Acute toxicological impact of nano- and submicro-scaled zinc oxide powder on healthy adult mice. *Bioinformatics and Biomedical Engineering 4th International Conference* 2010, (1–4).
- [194] Wang B., Feng W. Y., Wang M., Wang T. C., Gu Y. Q., Zhu M. T., Ouyang H., Shi J. W., Zhang F., Zhao Y. L., Chai Z. F., Wang H. F., Wang J., Acute toxicological impact of nano- and submicro-scaled zinc oxide powder on healthy adult mice. *Journal of Nanoparticle Research*, 10(2) (2008): 263–276.



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